

*2014 JSSX Award for Young Scientists (Oct 22, 2014)*



Molecular pharmacokinetic mechanisms of  
oxidative stress-induced tissue damage in  
chronic kidney disease for medical development  
and therapeutic application

慢性腎臓病における酸化ストレス臓器障害の  
分子動態学的解明と医療への展開

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# Chronic kidney disease: CKD

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- The prevalence of CKD in Japan is over 13 million patients, which is 13% among the Japanese general adults population.
- The number of patients with hemodialysis is exceeded 300,000.
- Dialysis costs more than 150 billion yen/year.
- CKD is a major risk factor for cardiovascular disease (CVD), which is a major cause of death in CKD patients.

Developing new therapeutic approaches for CKD is urgently necessary.

# Outline

## Therapeutic approaches to progression of CKD and CVD

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### 1. Molecular mechanism of uremic toxin-induced tubular and vascular damage

- *p*-Cresyl sulfate induced-renal tubular and vascular cell damages
- CMPF induced-cell damage *via* generation of a radical intermediate

CKD/Uremic toxicity and CVD

### 2. Biomarker identification

- Monitoring of cysteinylated albumin by using ESI-TOFMS

### 3. Therapeutic development

- Novel anti-oxidative/anti-inflammatory agent: Albumin-thioredoxin fusion protein

# Uremic toxins

Accumulated substances that interact negatively with biologic functions in CKD condition.

88 molecules were identified.

## 1. Low molecular weight molecules: 40 molecules

Guanidino succinate, Phenylacetic acid, Uric acid, Neopterin etc.

## 2. Middle molecules: 23 molecules

FGF23, PTH,  $\beta_2$ -MG, TNF- $\alpha$  etc.

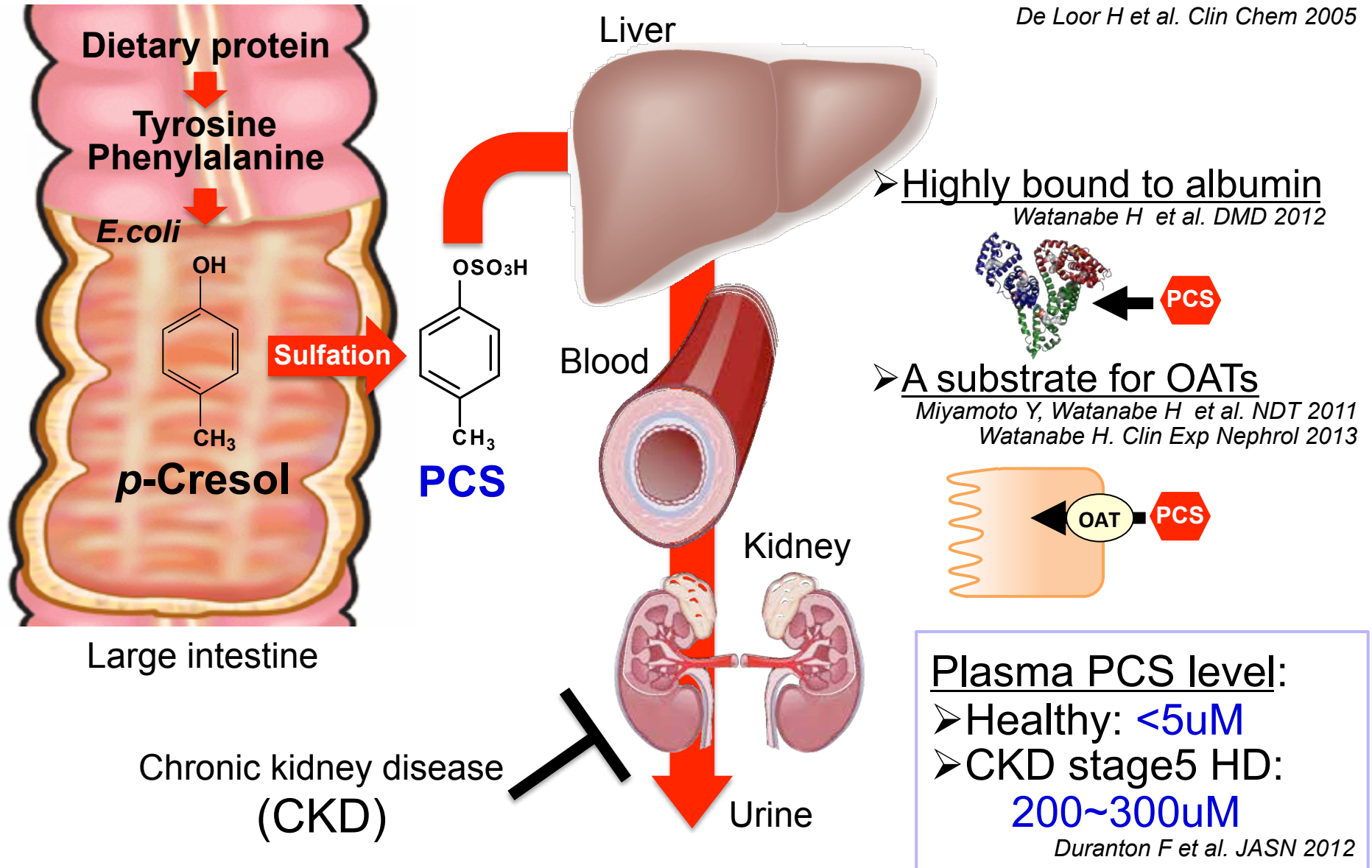
## 3. Protein-bound solutes: 25 molecules

p-Cresyl sulfate, CMPF, Indoxyl sulfate, etc.

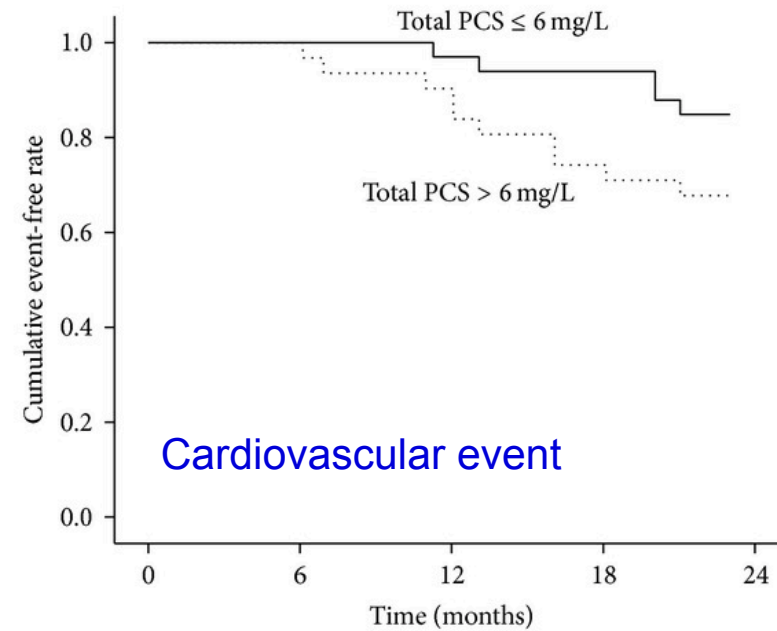
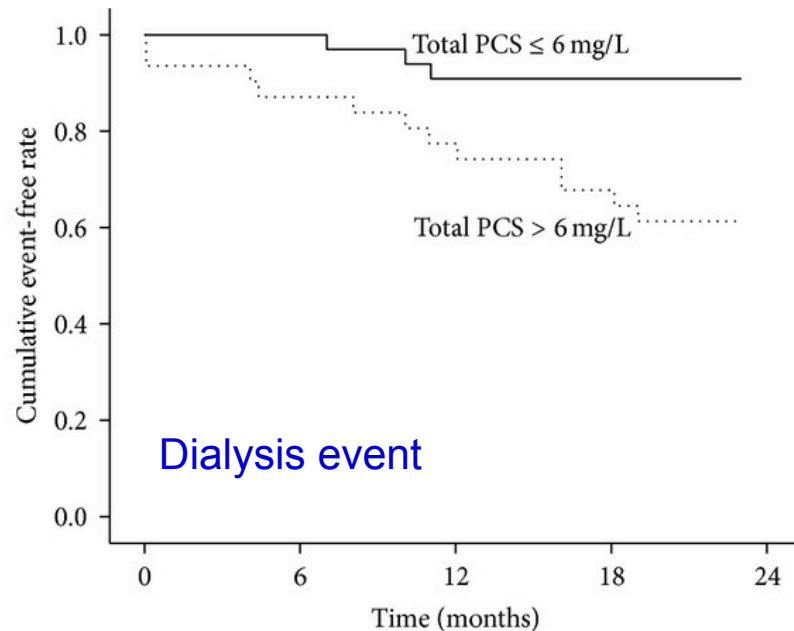
**Insufficient removing by dialysis**

# p-Cresyl sulfate: PCS

Martinez AW et al. CJASN 2005  
De Loor H et al. Clin Chem 2005



# Plasma PCS levels associated with CKD progression, cardiovascular mortality



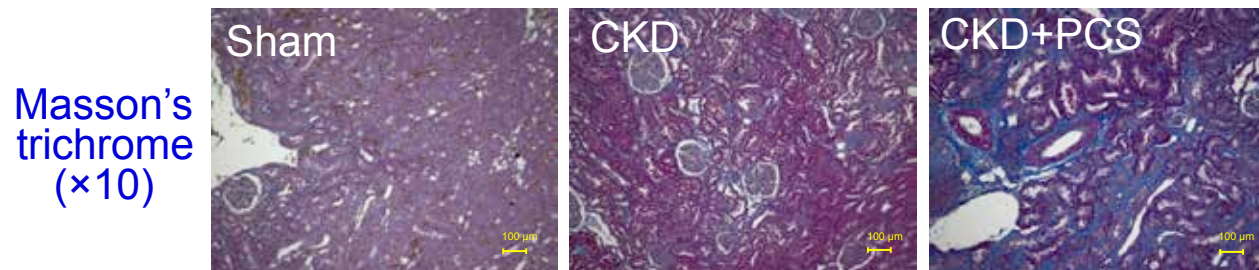
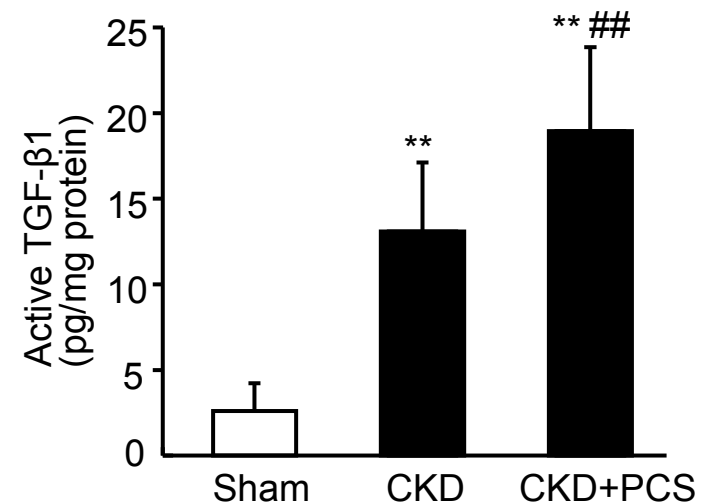
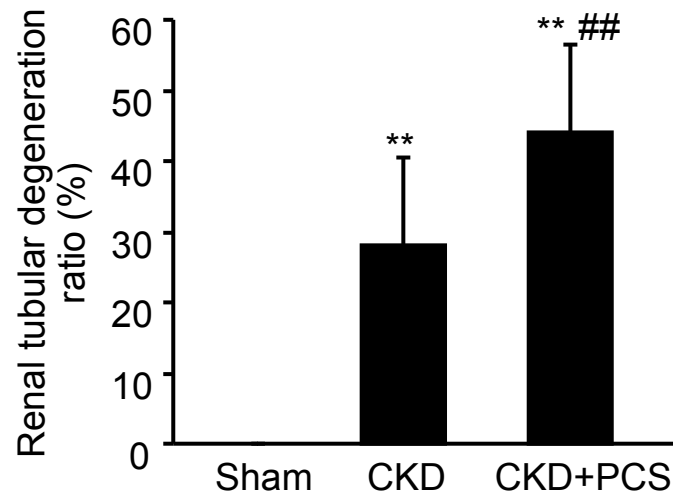
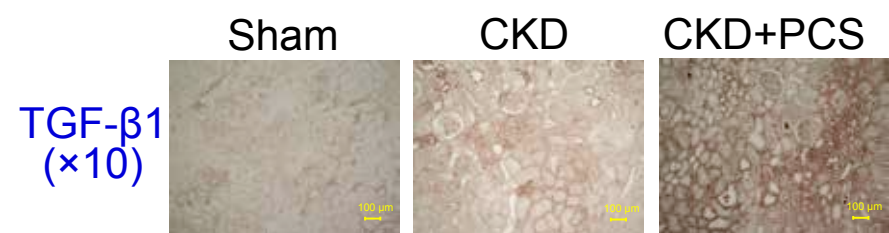
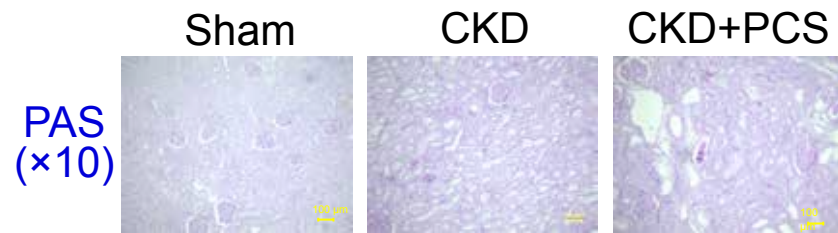
Clinical studies showed higher plasma PCS levels associated with 1.dialysis event and 2.cardiovascular event.

However, the biological functions of PCS in kidney/vascular system, and the mechanism of its action were largely unknown.



*Lin CJ et al. Biomed Res Int. (2014)*  
*Cheng JL et al. Atherosclerosis (2012)*  
*Wu I et al. NDT (2011&2012)*  
*Liabeuf S et al. NDT (2010)*  
*Meijers BK et al. CJASN (2010)*  
*Bammens B et al. Kidney Int. (2006)*

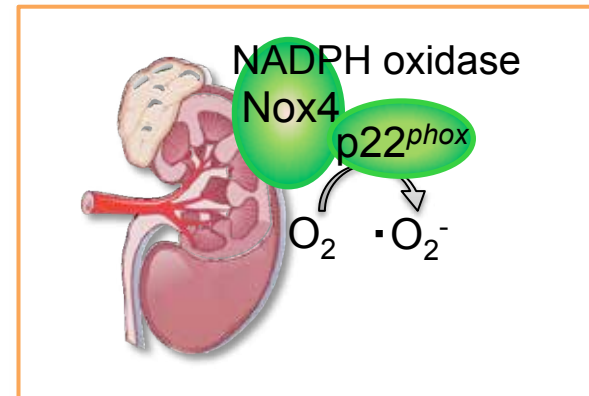
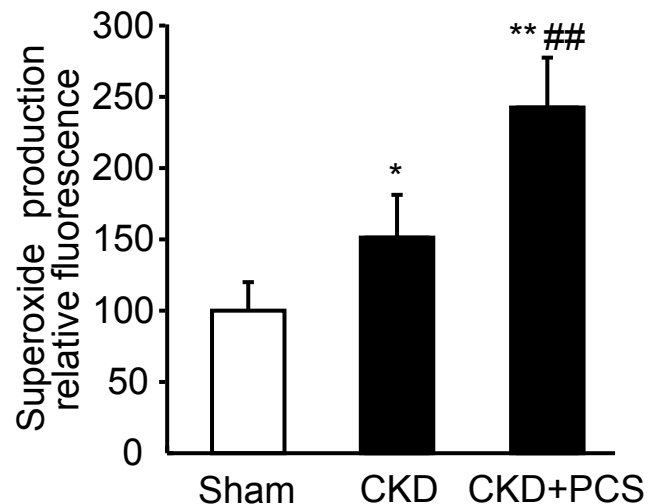
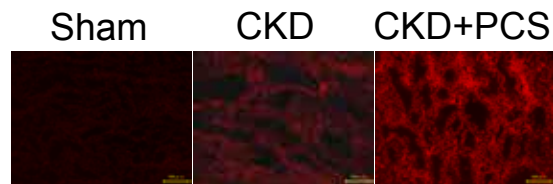
# Histological evaluation of kidney from PCS-overloaded CKD rats



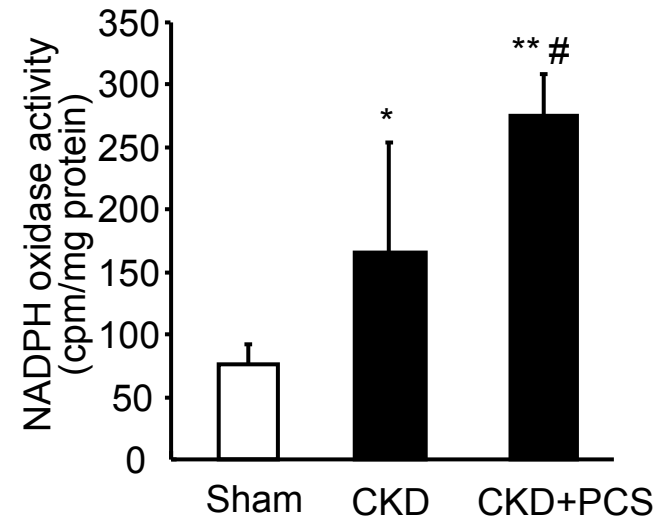
PCS-overload increased tubular damage, especially, tubular degeneration, TGF- $\beta 1$  expression and renal fibrosis.

# Superoxide production and NADPH oxidase activity in kidney from PCS-overloaded CKD rats

## Superoxide production (DHE stain)



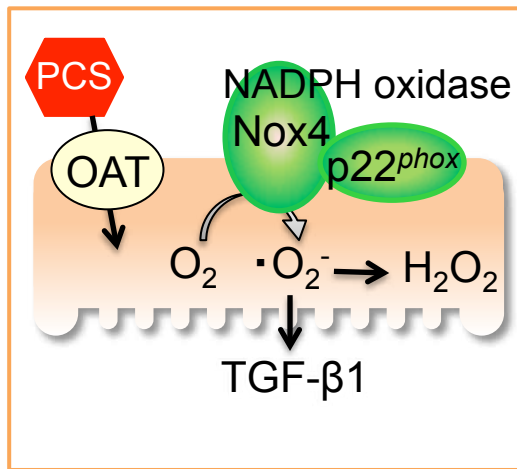
## NADPH oxidase activity



PCS-overload showed significant increase in superoxide production and NADPH oxidase activity.



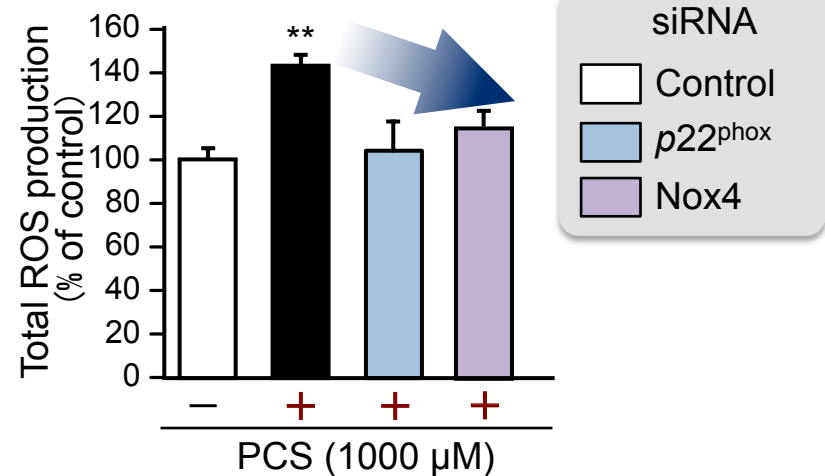
# siRNA for NADPH oxidase and probenecid suppressed PCS-induced ROS production and TGF- $\beta$ 1 expression (HK-2)



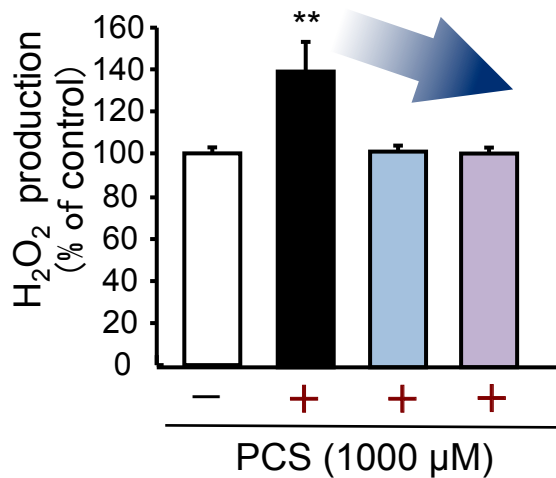
Nox4/p22<sup>phox</sup> knockdown



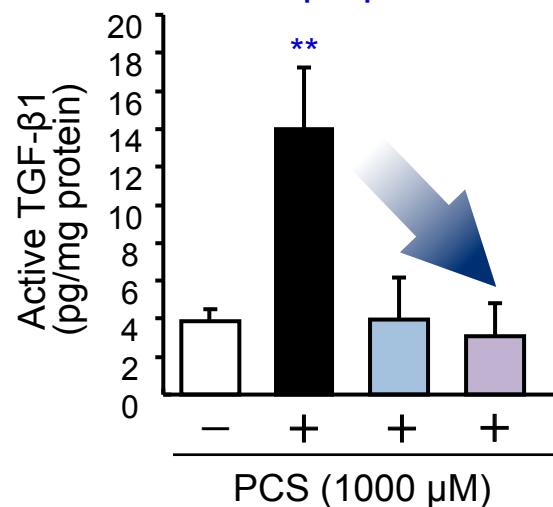
Total ROS production



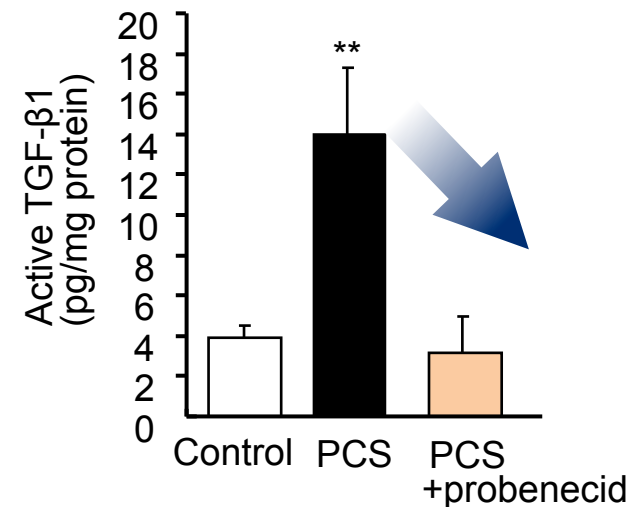
H<sub>2</sub>O<sub>2</sub> production



Active TGF- $\beta$ 1 protein



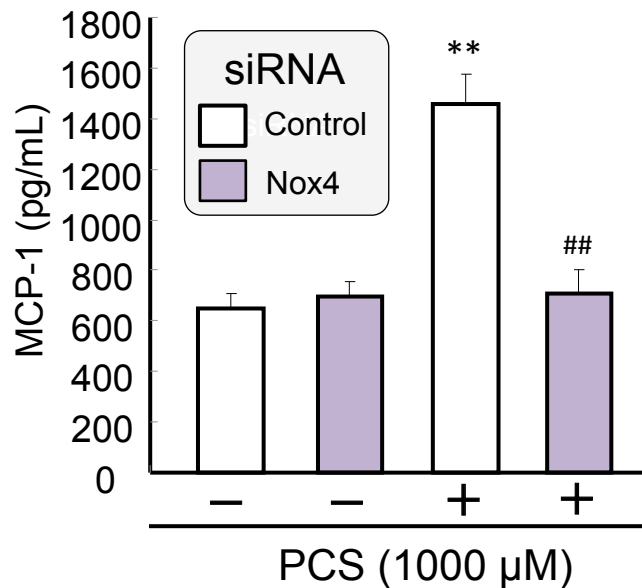
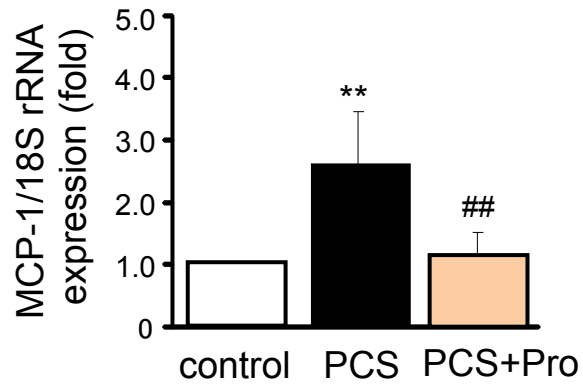
Active TGF- $\beta$ 1 protein



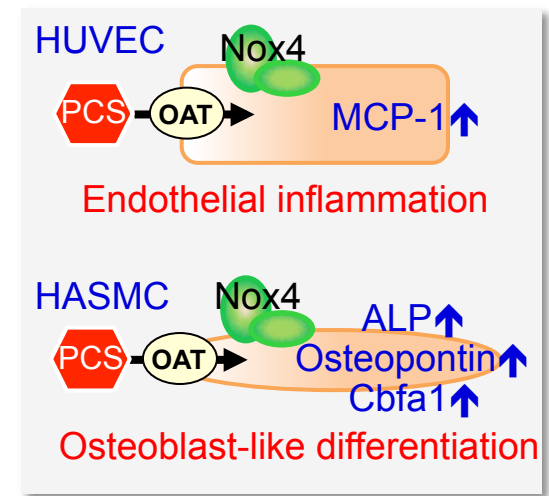
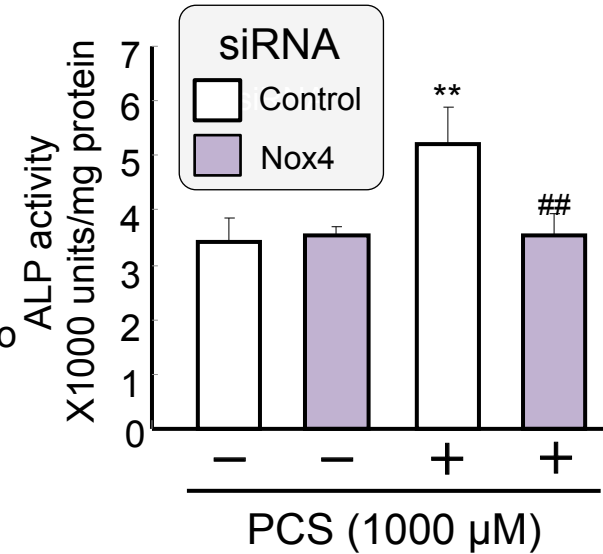
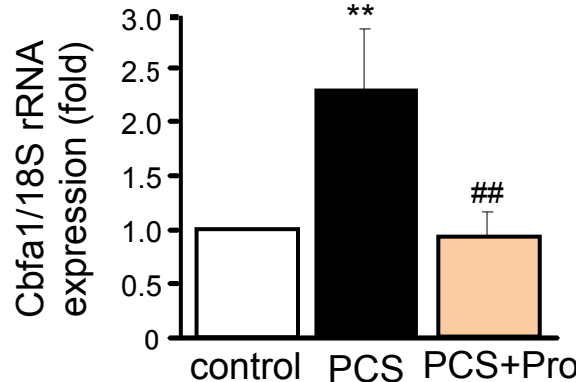
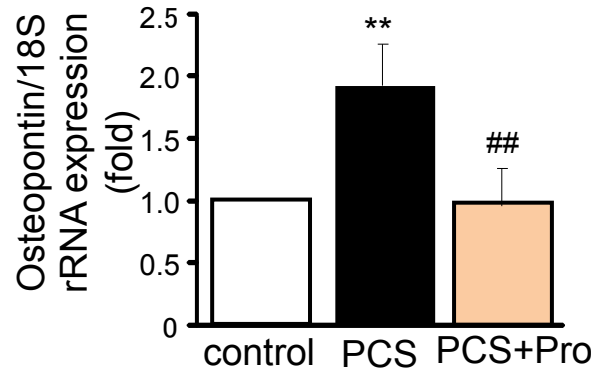
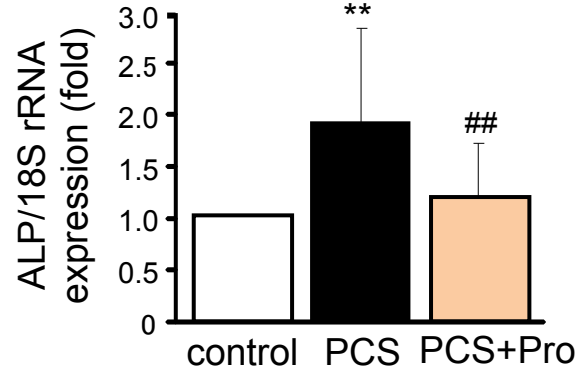
Cellular accumulation of PCS *via* OATs activates NADPH oxidase, which contributes to ROS production and increasing TGF- $\beta$ 1 expression.

# PCS increased MCP-1 expression in HUVEC and osteoblast-specific genes in HASMC

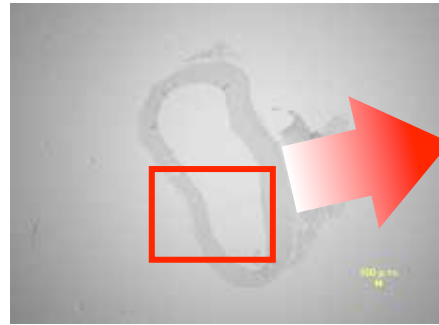
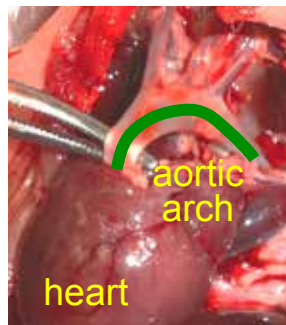
## Vascular endothelial cell (HUVEC)



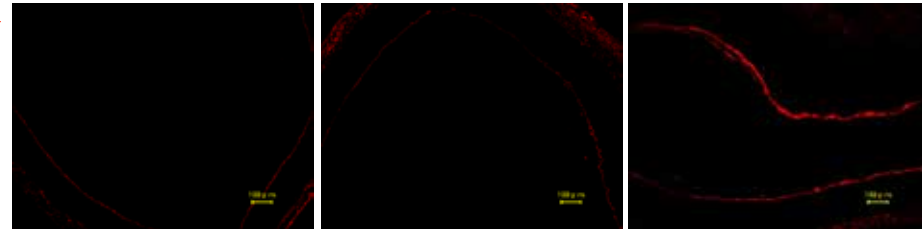
## Vascular smooth muscle cell (HASMC)



# PCS-overload leads to vascular damage in CKD rats



NADPH oxidase (Nox4) (x10)  
(aortic arch)



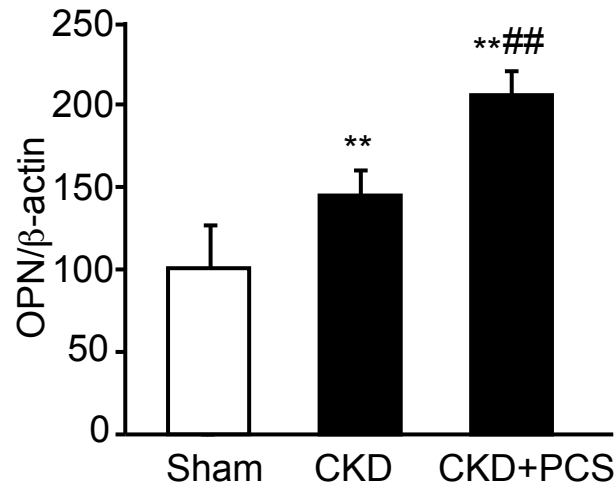
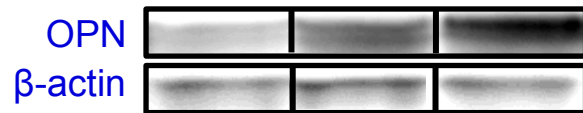
Sham

CKD

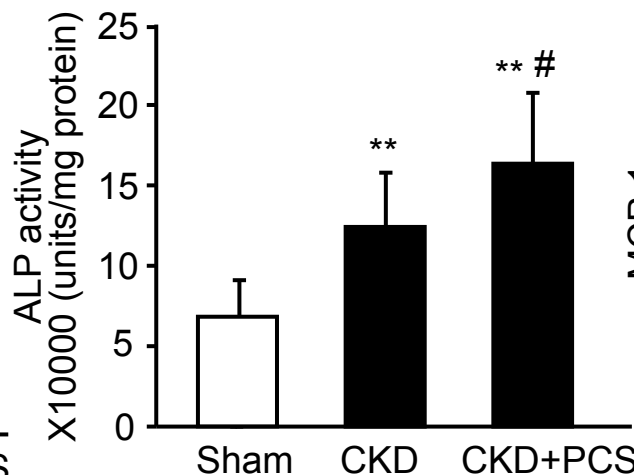
CKD+PCS

Osteopontin

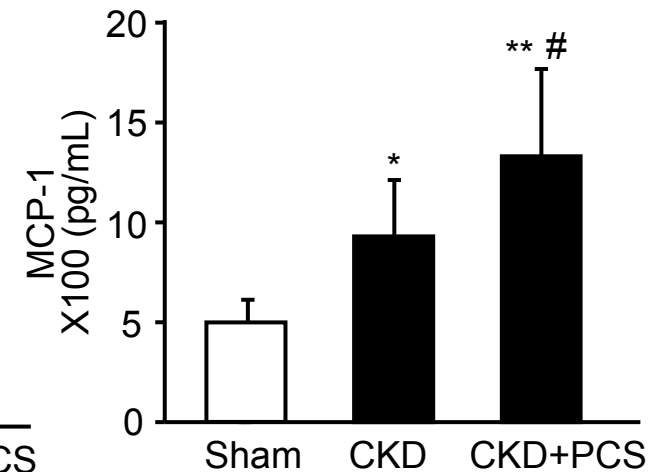
(aortic arch homogenate)



ALP activity  
(aortic arch homogenate)

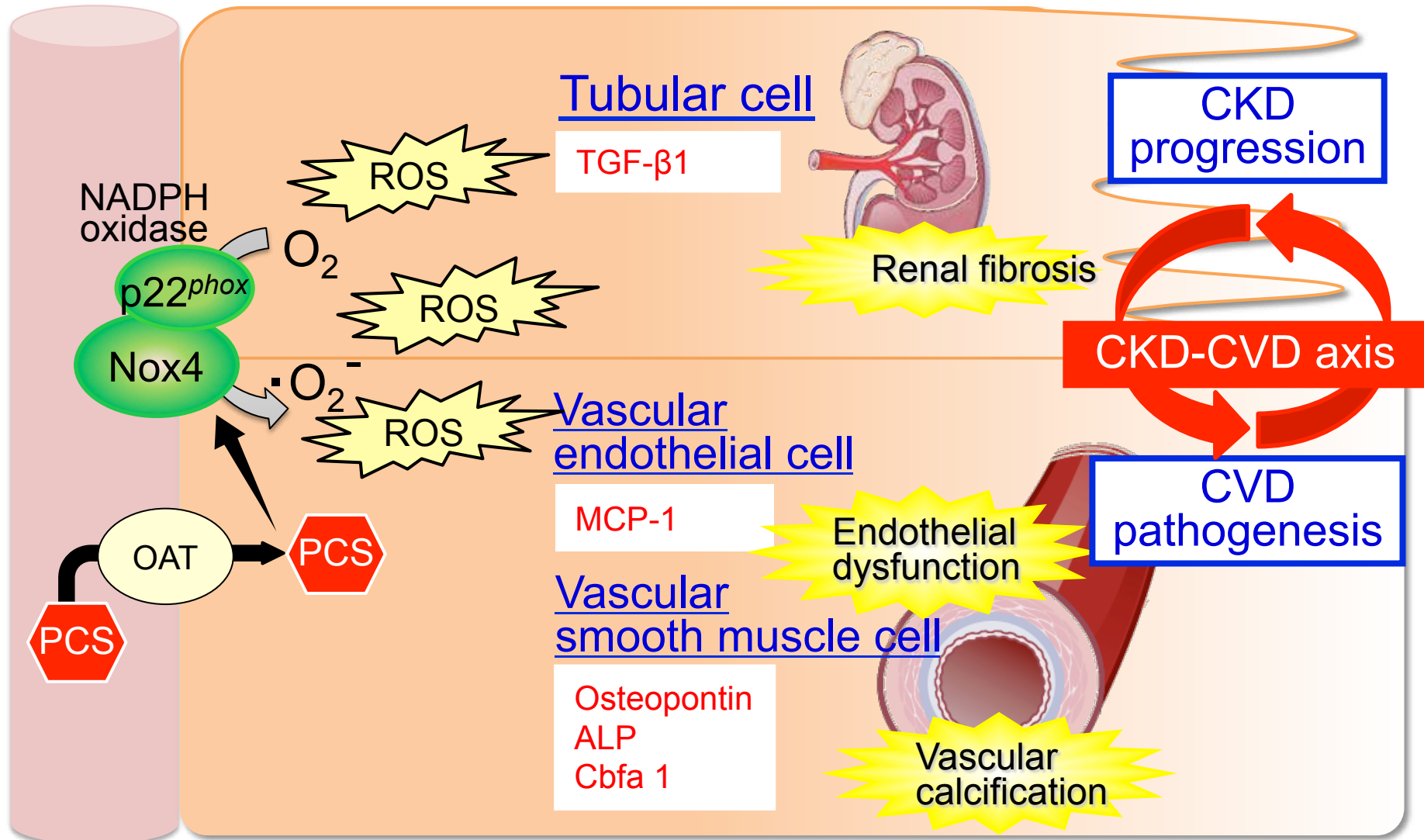


MCP-1 level  
(plasma)



In aortic arch, PCS-overload increases the expression of NADPH oxidase, osteopontin and ALP activity in addition to increasing plasma MCP-1 level.

# Intracellular accumulation of PCS *via* OATs could contribute to CKD-CVD axis

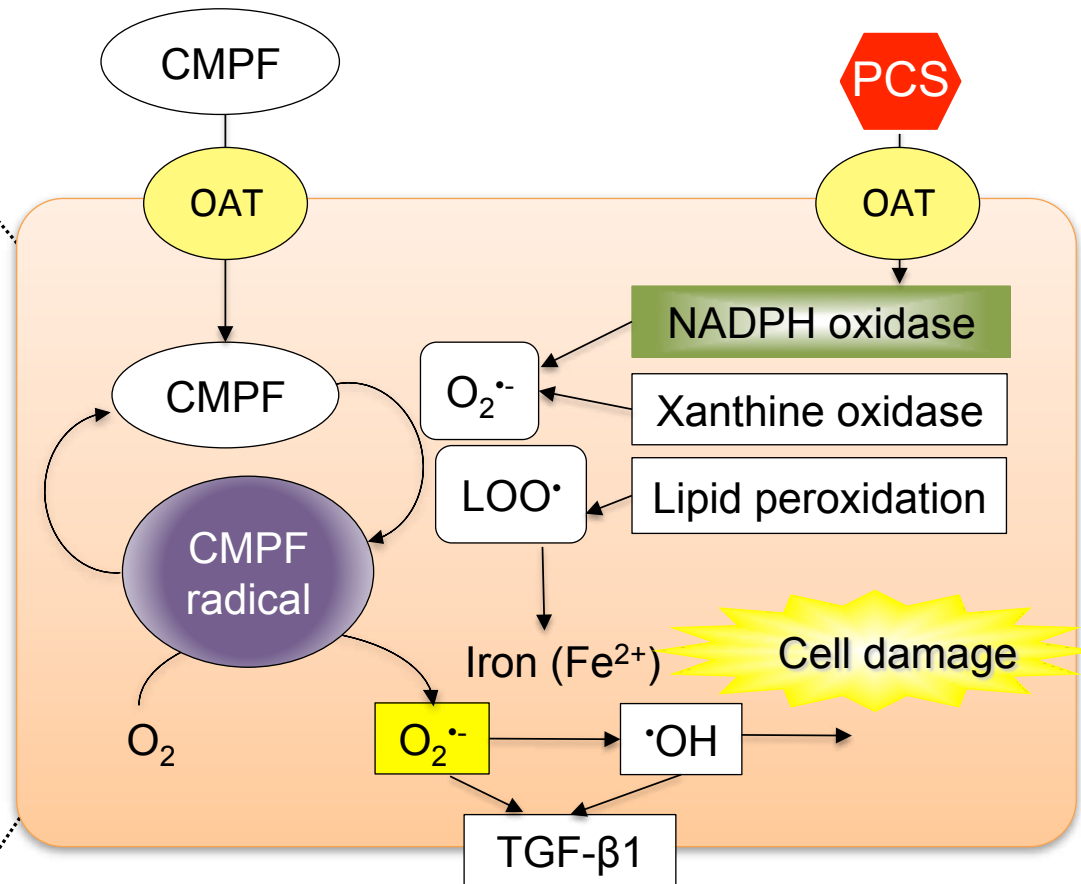
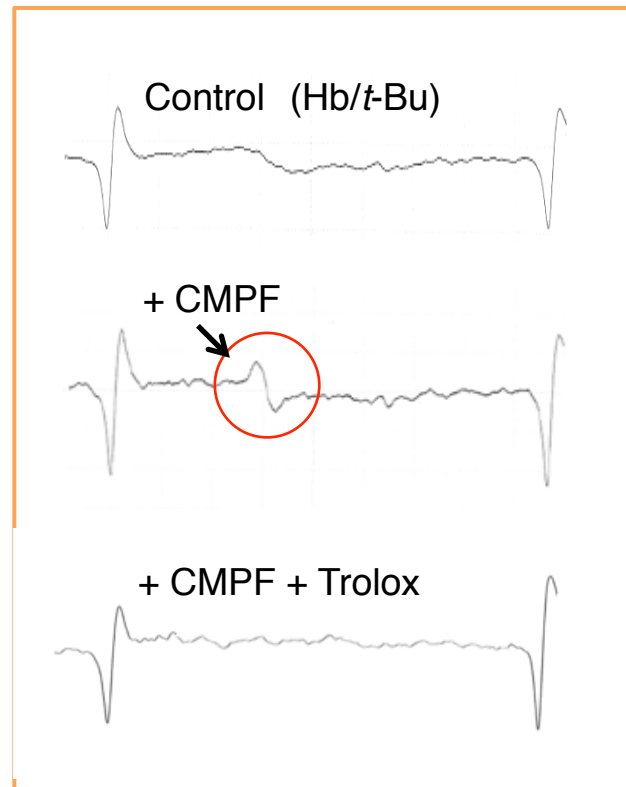
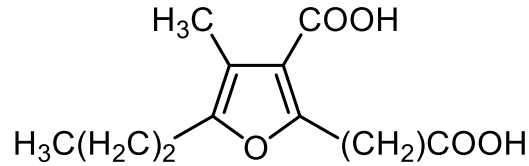


Watanabe H et al. *Kidney Int.* 2013

Watanabe H et al. *Pharmacol Res Perspect.* 2014

# CMPF induces cell damage *via* generation of a radical intermediate

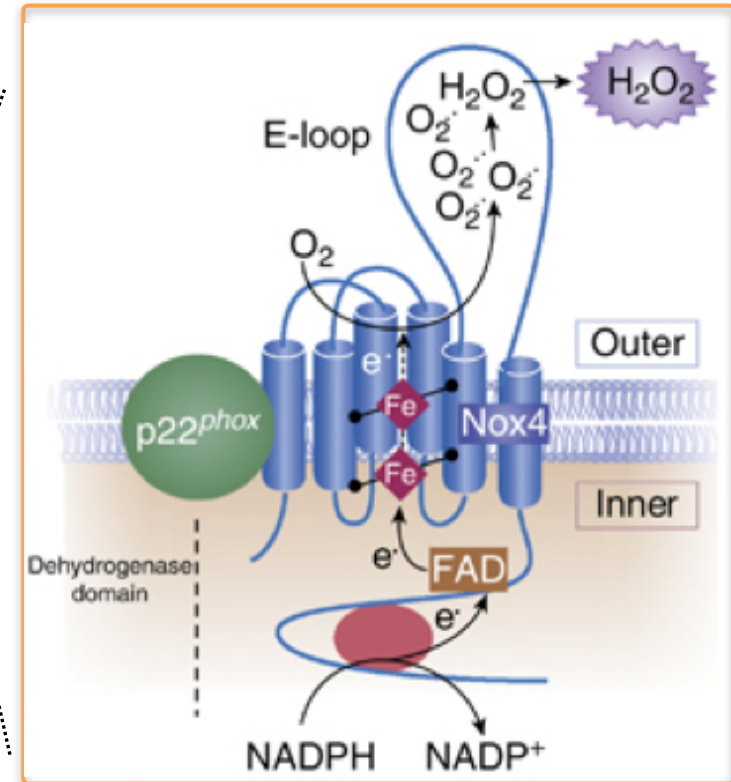
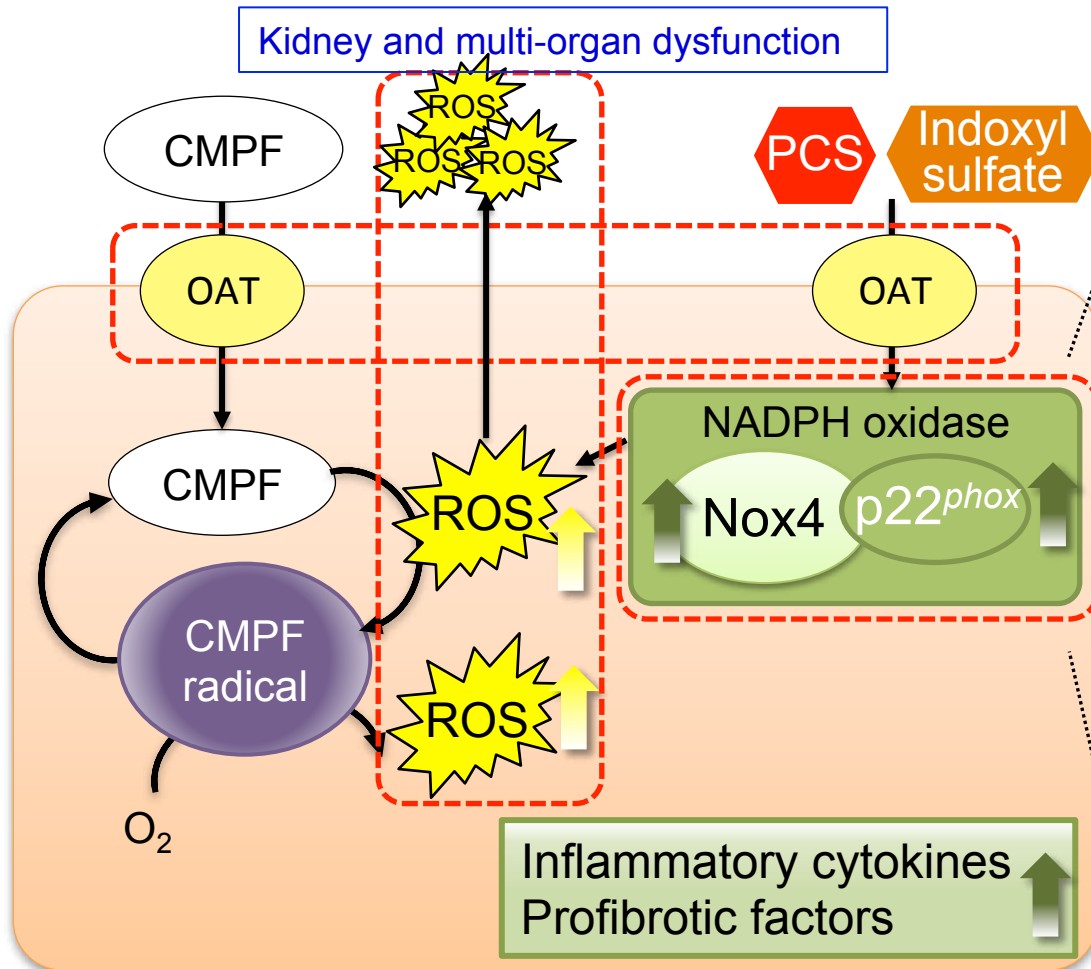
CMPF: 3-Carboxy-4-methyl-5-propyl-2-furanpropionate



CMPF interacts with superoxide anion radicals ( $O_2^{\cdot-}$ ) and peroxy radicals to produce CMPF radicals, which lead to cellular damage *via* overproduction of  $O_2^{\cdot-}$ .

# Potential therapeutic target for treatment of uremic toxicity associated to CKD

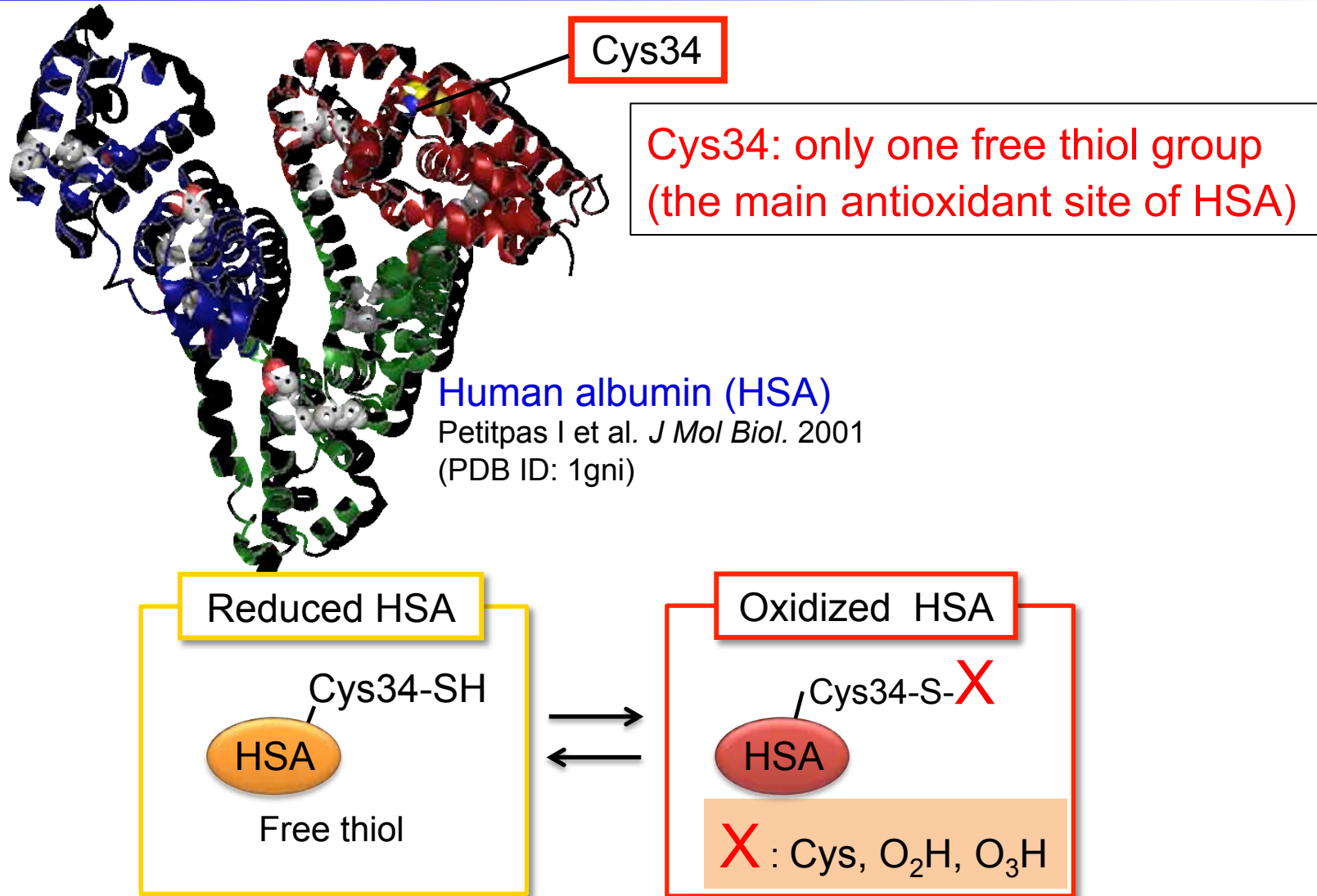
Commentary from Gorin Y. (Univ. Texas)  
*Kidney Int.* 2013



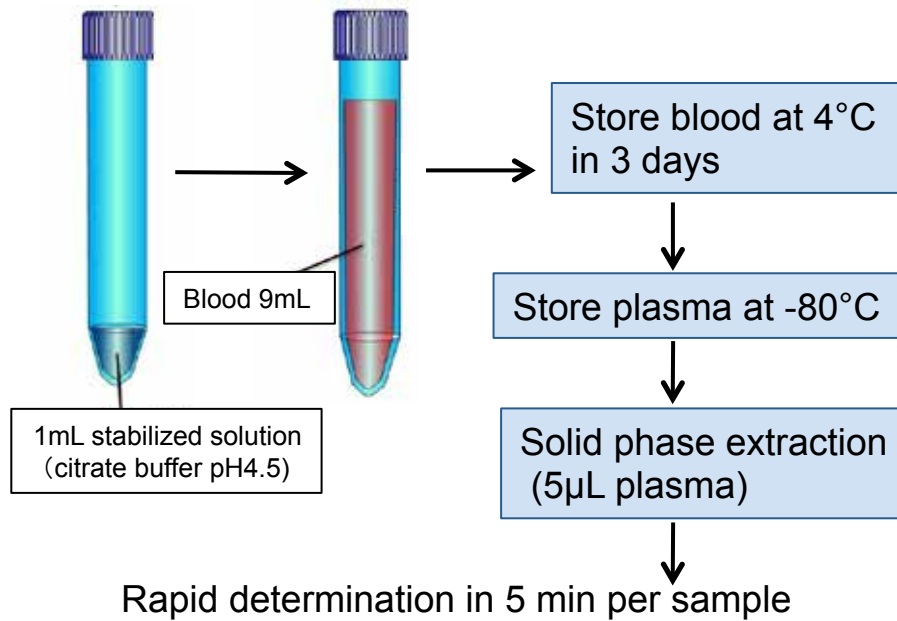
1. Cellular uptake pathway of uremic toxins, 2. NADPH oxidase, 3. Increased ROS

Our findings unraveled molecular mechanism of uremic toxin-induced progression of CKD and CVD.

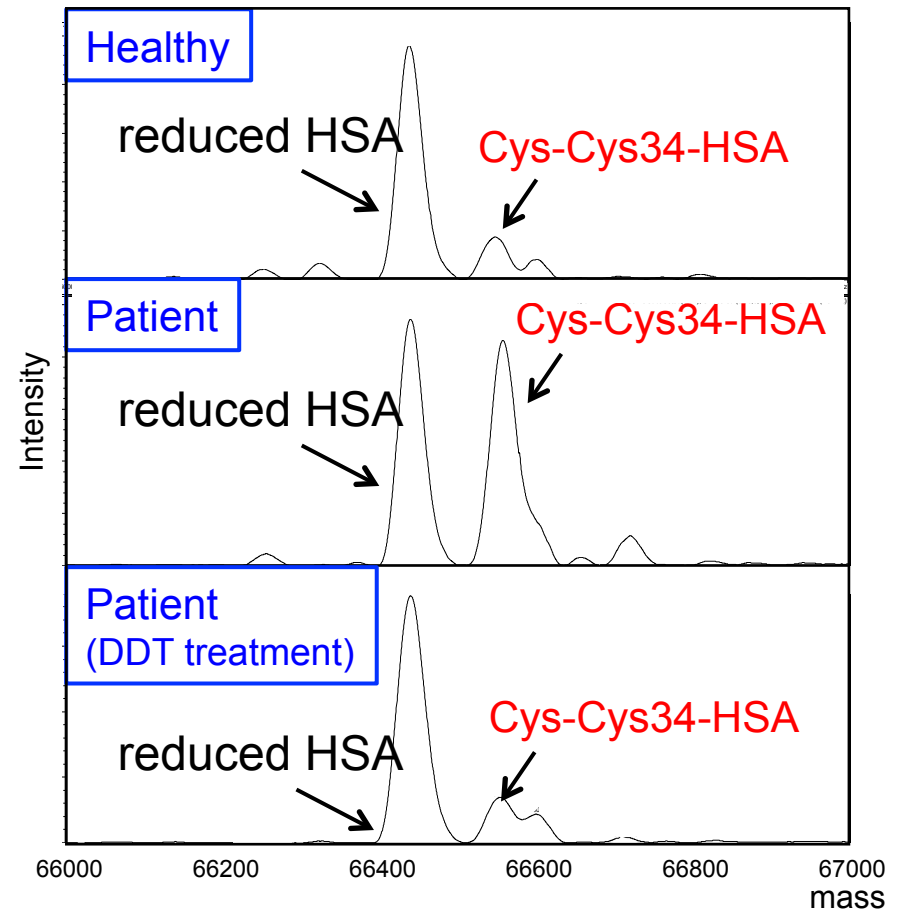
# New biomarker for CKD progression (Cys34-cysteinylated human albumin)



# Detection of redox change of Cys34 by using ESI-TOFMS



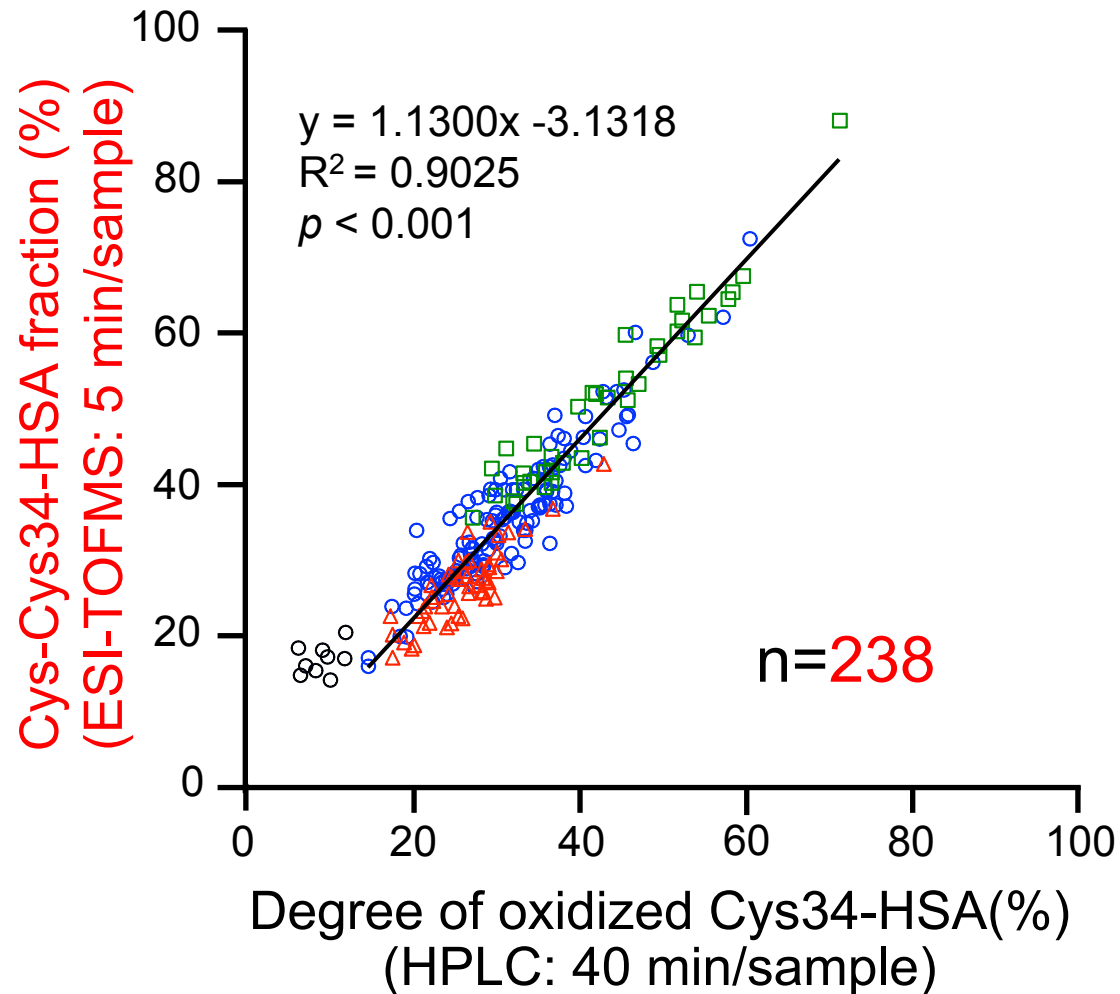
ESI-TOFMS spectra of HSA from healthy and patients



Major oxidized form is a disulfide-bonded Cys at Cys34 of HSA (Cys-Cys34-HSA)



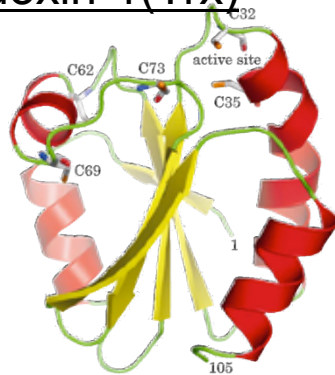
# Cys-Cys34-HSA fraction measured by using ESI-TOFMS is a new biomarker for oxidative stress



This system is a suitable analytical method for rapid and sensitive clinical laboratory analysis.

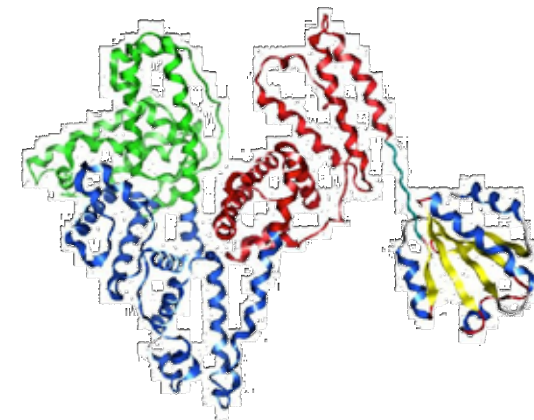
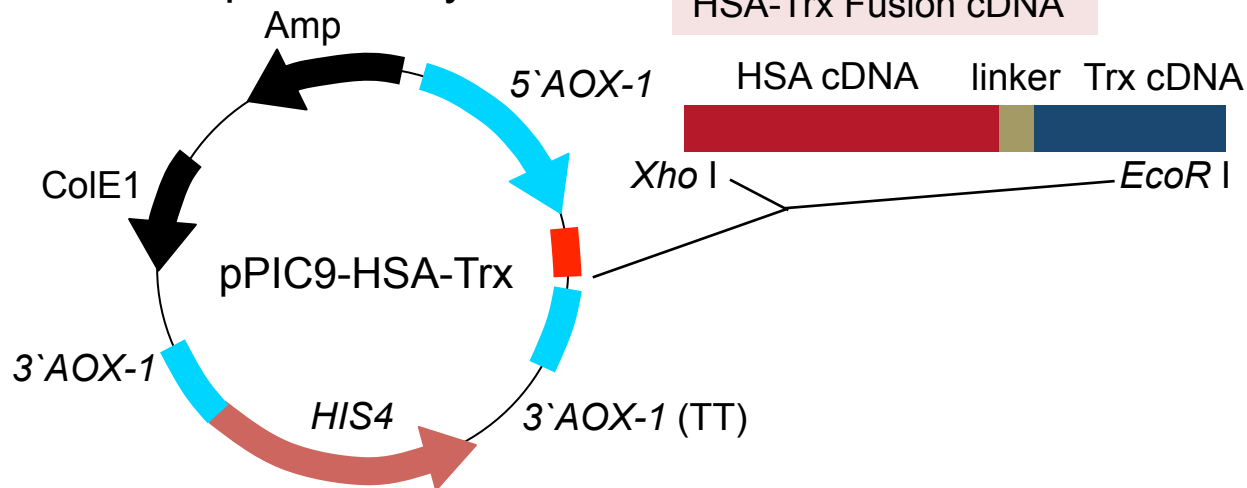
# Novel anti-oxidative and anti-inflammatory agent albumin (HSA)-thioredoxin (Trx) fusion

## Thioredoxin-1(Trx)



- ✓ anti-oxidative action
- ✓ anti-inflammatory action
- ✓ anti-apoptotic action
- ✓ plasma half-life ( $t_{1/2}$ ) in mouse: 1 hr

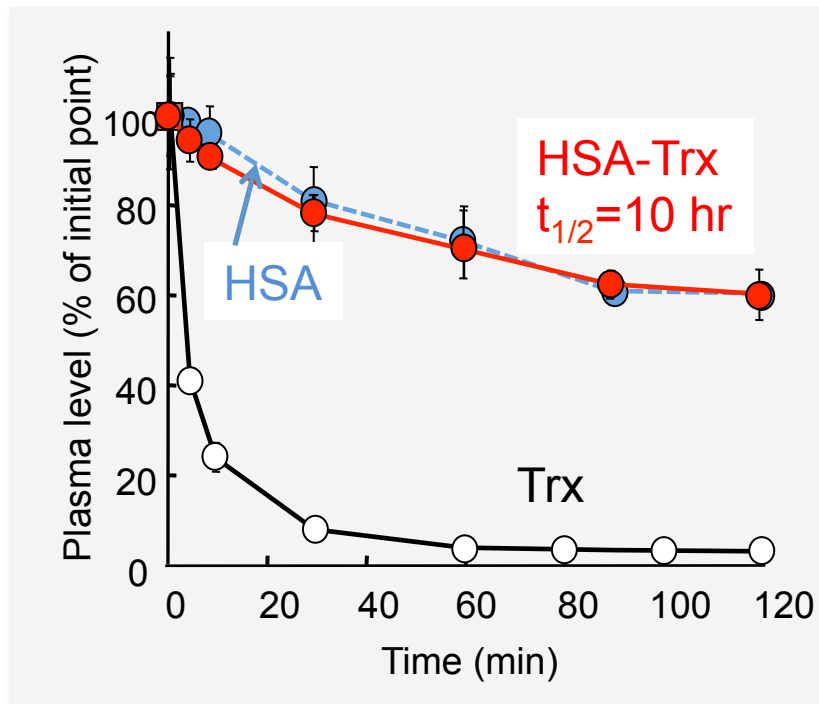
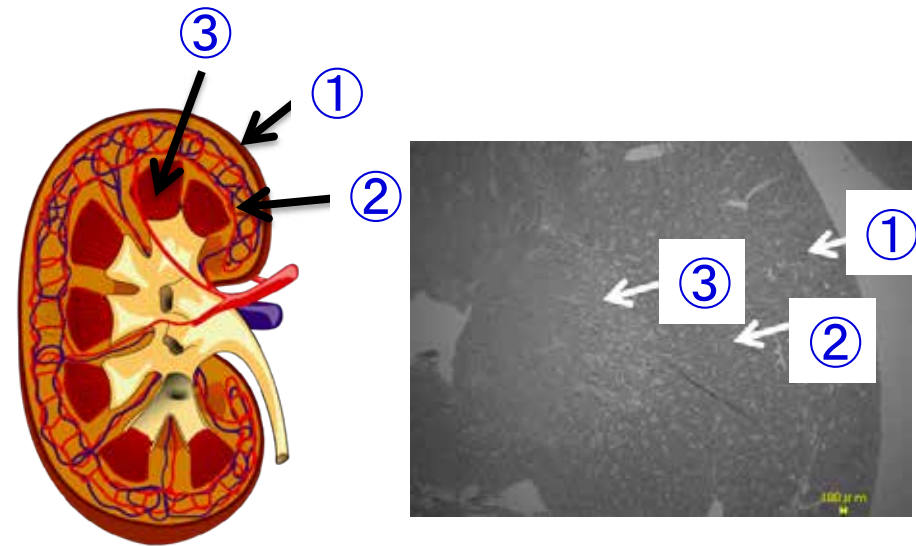
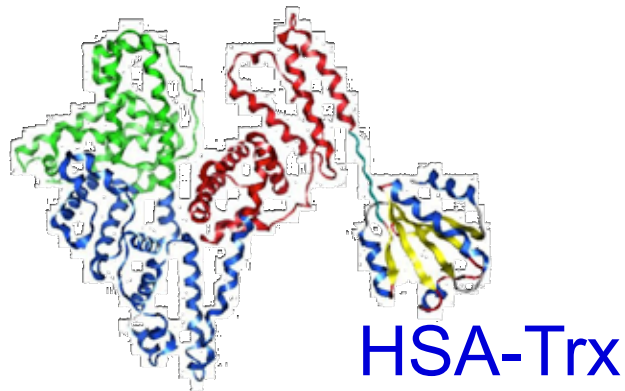
## Pichia expression system



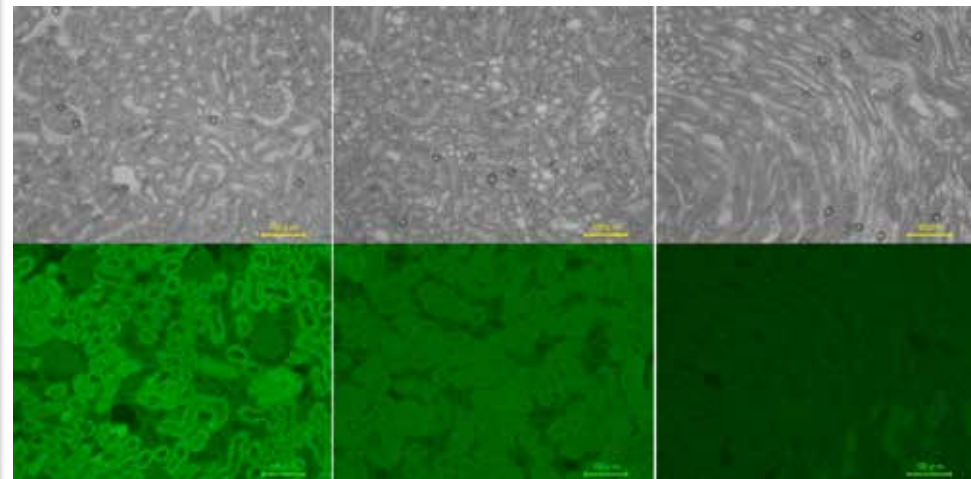
HSA-Trx  
(image view)

Ikuta S et al. *J Control Release* 2010, Furukawa M et al. *J Control Release* 2011, Tanaka R et al. *JPET* 2013, Kodama A et al. *Kidney Int.* 2013, Kodama A et al. *BBA.* 2014, Tanaka R et al. *Mol Pharm.* 2014

# Plasma level-time curve and renal localization of HSA-Trx



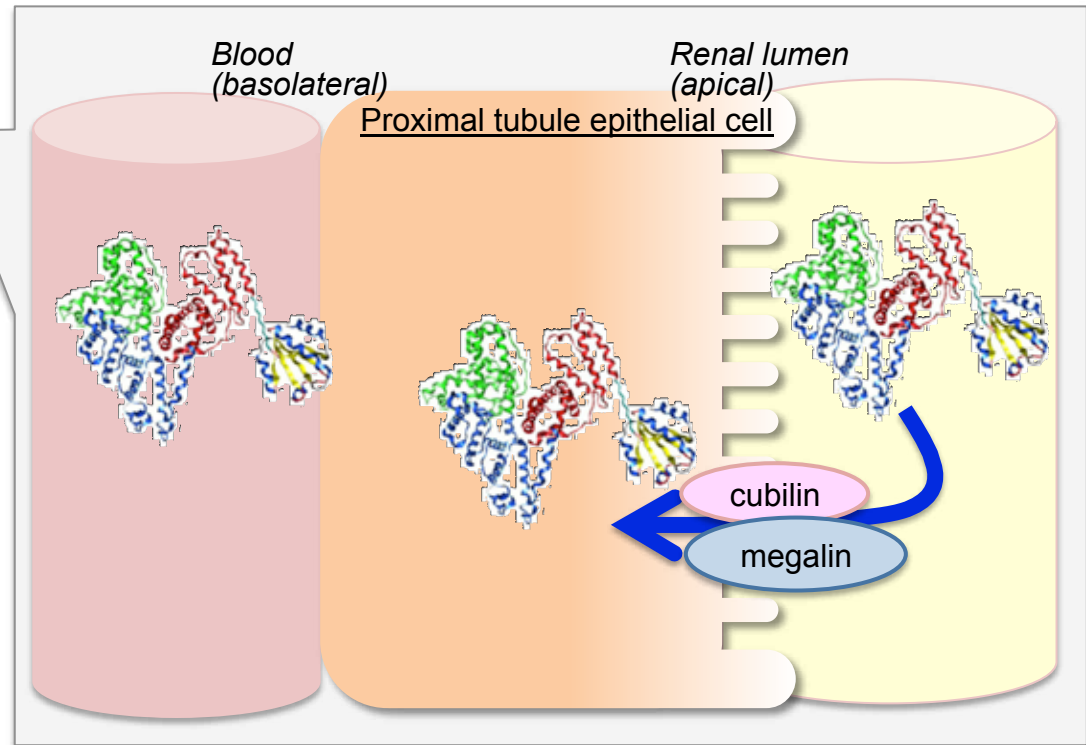
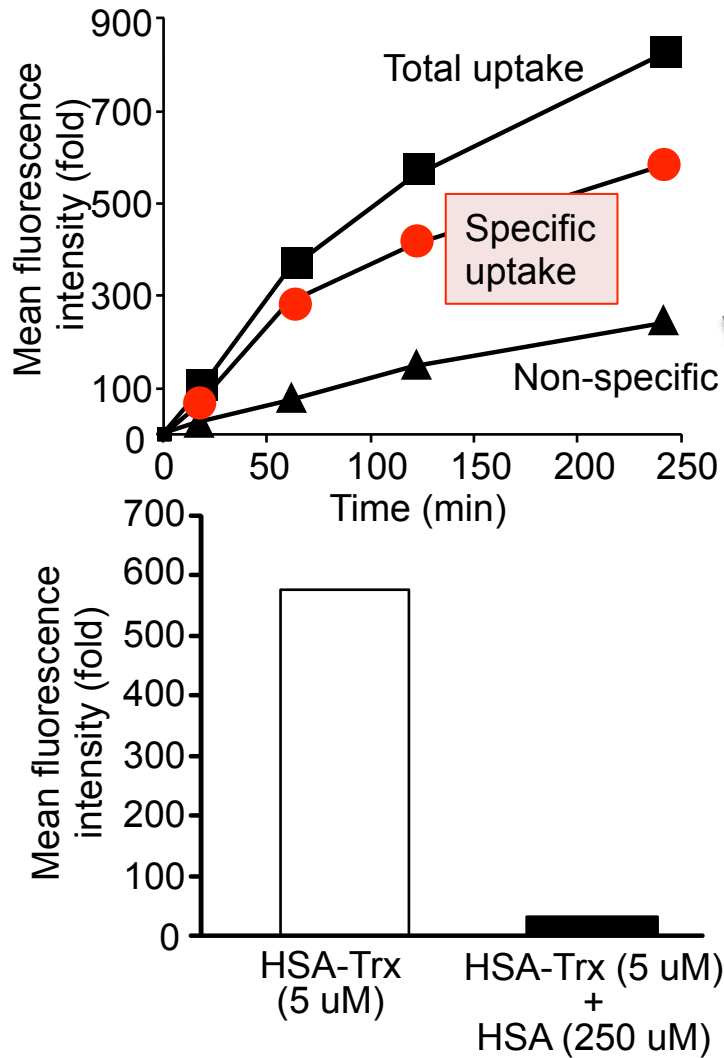
①Cortex      ②Cortico-medullary boundary zone      ③Medulla



Ikuta S et al. *J Control Release* 2010,  
Furukawa M et al. *J Control Release* 2011

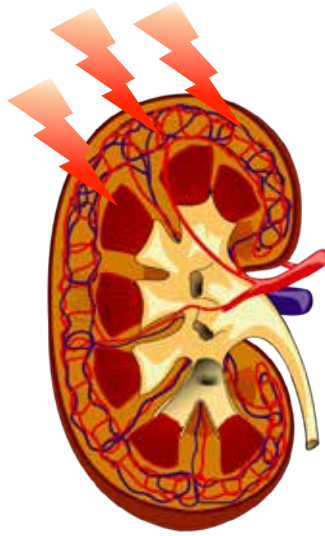
Kodama A et al. *BBA*. 2014

# Uptake of HSA-Trx by human proximal tubular cell (HK-2) via megalin/cubilin-mediated endocytic pathway

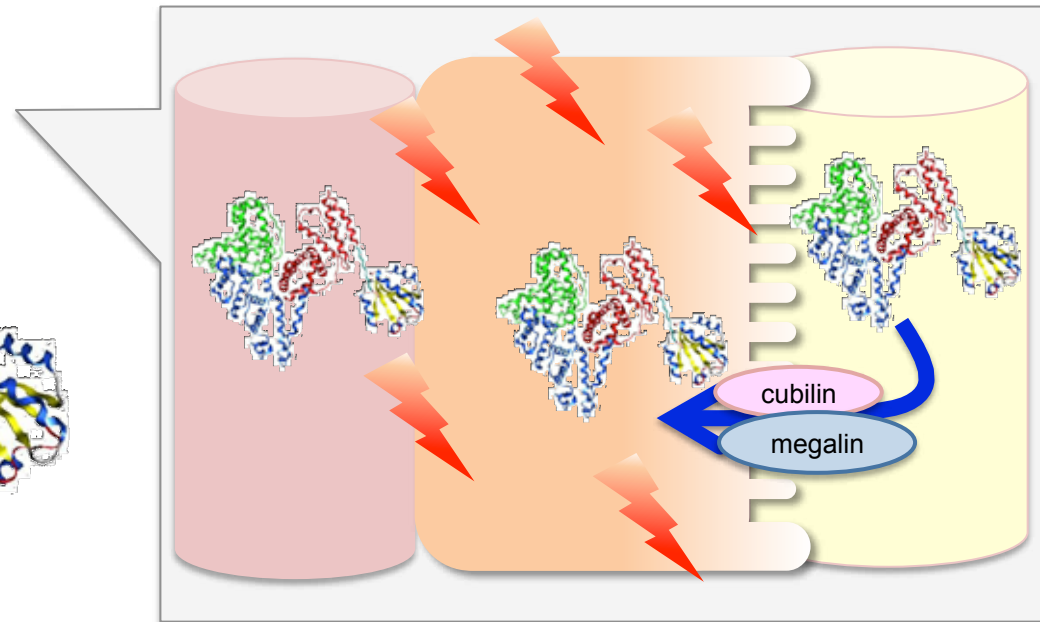
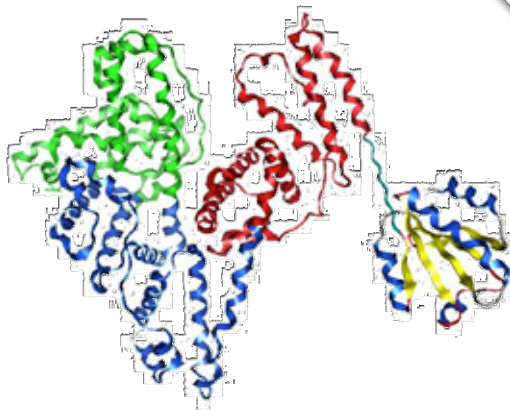


HSA-Trx can act not only in extracellular compartment but also inside the proximal tubular cells.

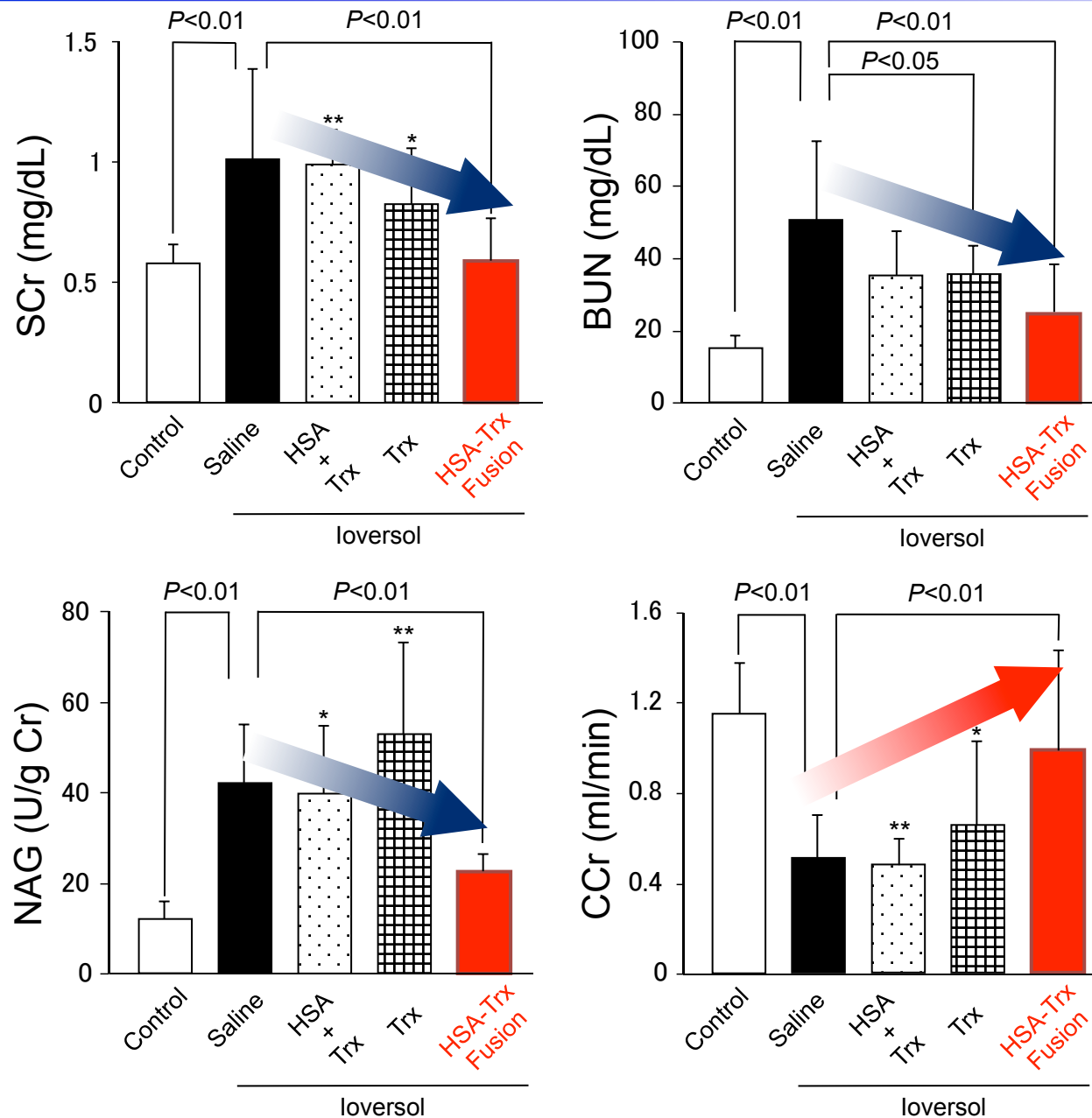
# Renoprotective effect of HSA-Trx on nephropathy model animals



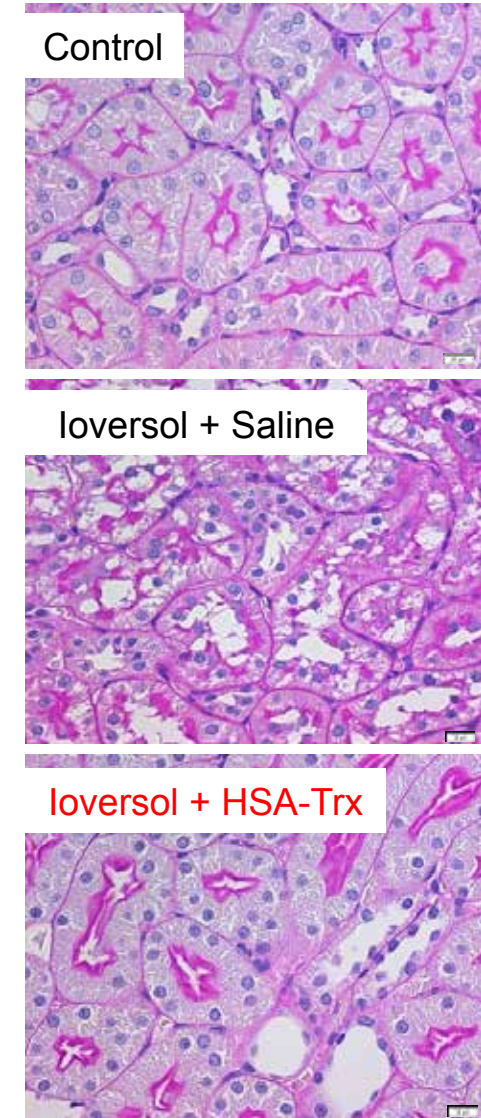
1. Contrast-induced nephropathy
2. Cisplatin-induced nephropathy
3. Rhabdomyolysis nephropathy



# Renoprotective effect of HSA-Trx on contrast-induced nephropathy

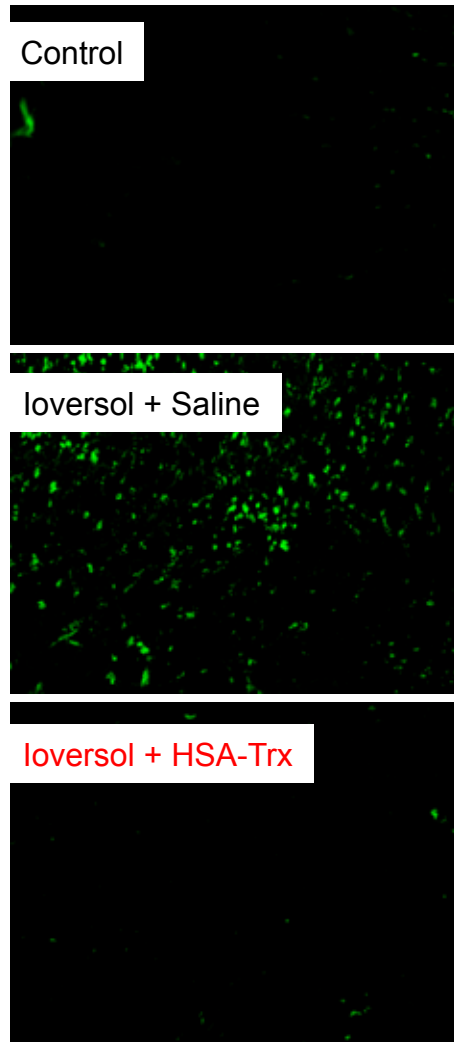


## PAS staining

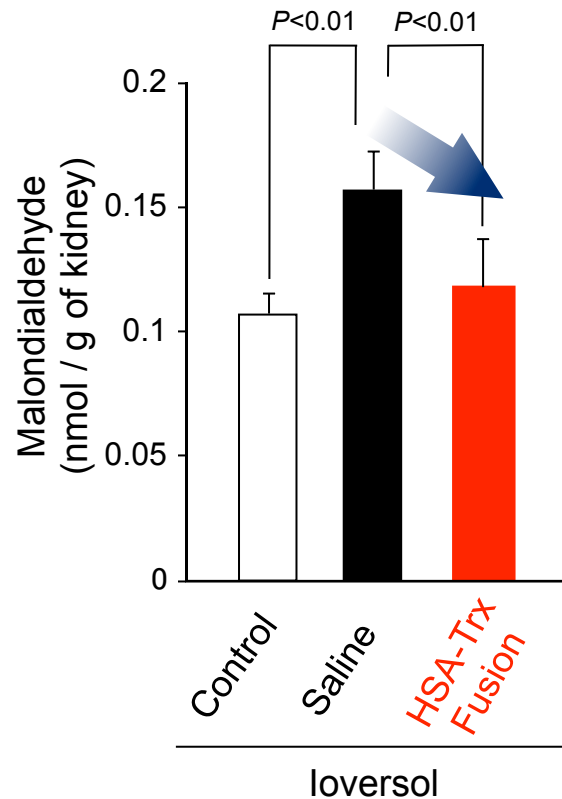


# The redox effect and anti-apoptotic effect of HSA-Trx on contrast-induced nephropathy

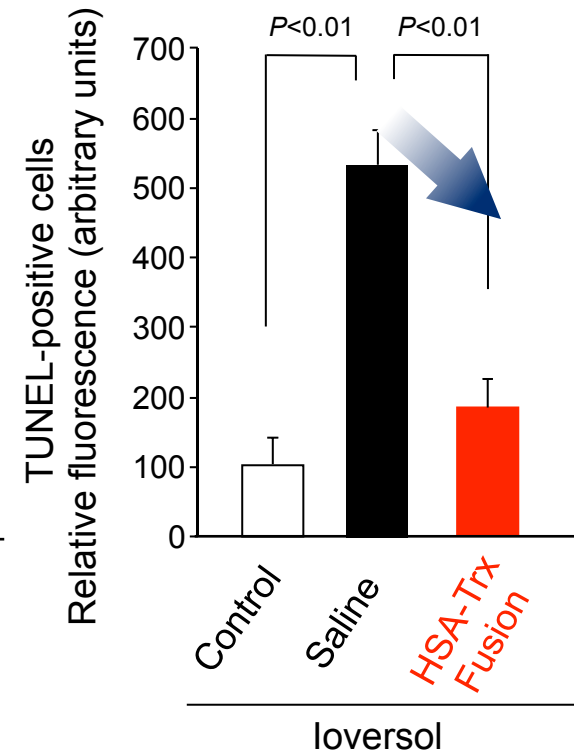
### 8-OHdG (Oxidative DNA damage)



### Malondialdehyde (Oxidative lipid damage)

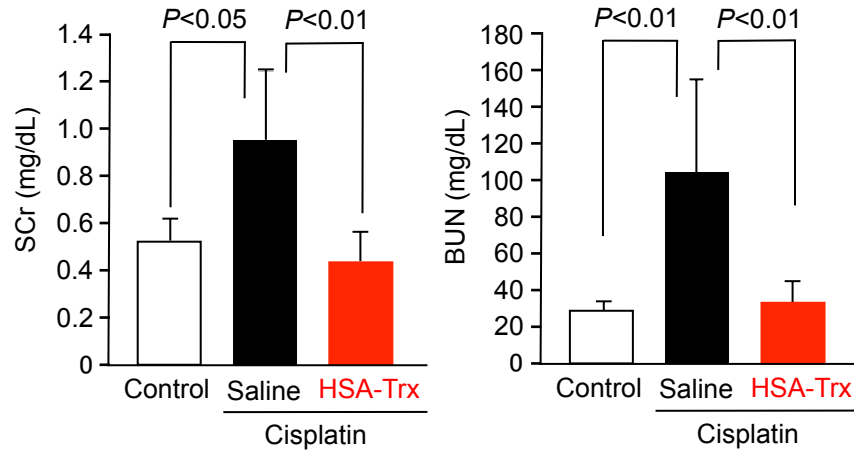


### TUNEL positive cells

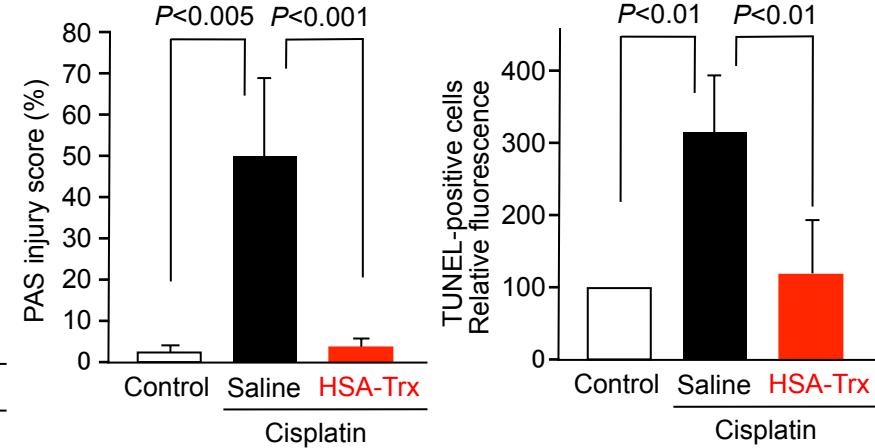


# HSA-Trx prevents cisplatin-induced nephropathy

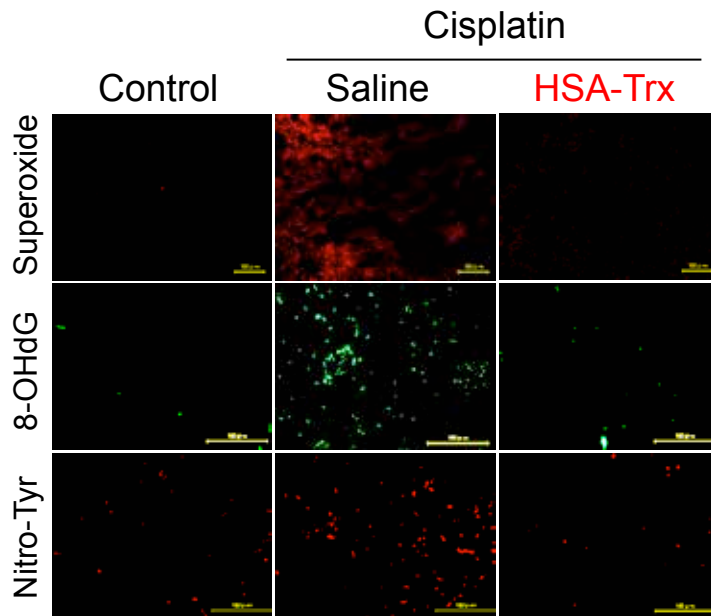
## Renal injury



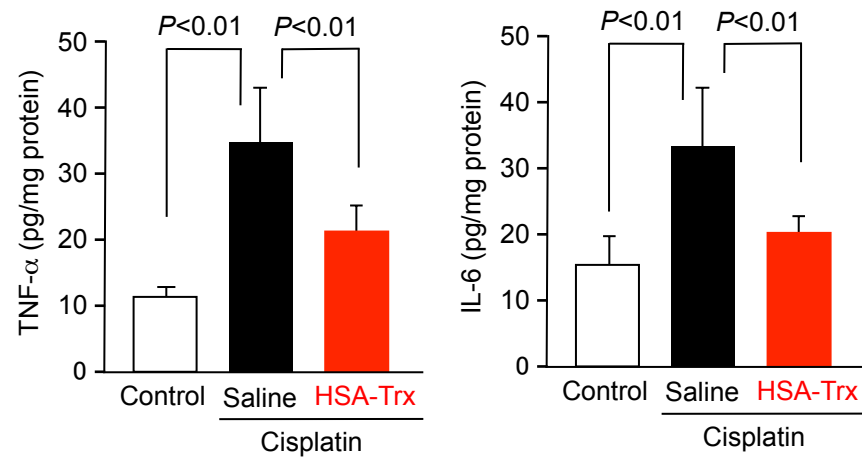
## Renal Apoptosis



## Renal oxidative stress



## Renal inflammation





# Summary

## Therapeutic approaches to progression of CKD and CVD

### 1. Molecular mechanism of uremic toxin-induced tubular and vascular damage

Miyamoto Y et al. *FEBS Lett.* 2010

Miyamoto Y et al. *Nephrol Dial Transplant.* 2011

Miyamoto Y et al. *Ther Apher Dial.* 2011

Watanabe H et al. *J Pharm Sci.* 2011

Miyamoto Y et al. *Biochem Pharmacol.* 2012

Watanabe H et al. *Drug Metab Dispos.* 2012

Tanaka H et al. *Bone* 2013

Watanabe H et al. *Kidney Int.* 2013

Watanabe H et al. *Pharmacol Res Perspect.* 2014

CKD/Uremic toxicity and CVD  
(Oxidative stress)

### 2. Biomarker identification

Nagumo K et al. *PIOS ONE* 2014

### 3. Therapeutic development

Tanaka R et al. *JPET* 2013

Kodama A et al. *Kidney Int.* 2013

Kodama A et al. *BBA* 2014

Tanaka R et al. *Mol Pharm.* 2014

The findings provide new insight into the pathogenesis of CKD and lead to new therapeutic approaches to diagnosis and treatment of CKD.

# Acknowledgments

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(National Cancer Center Research Inst.)

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## Dartmouth Medical School, USA

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## Tokai University

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## All members of Biopharmaceutics laboratory

