

***In vitro* ADME studies aiming at characterization of molecular mechanisms behind drug pharmacological or toxicological actions**

(ヒト薬効・毒性発現機序解明およびその評価能向上を目指した*in vitro*薬物動態研究)

Tomomi Furihata



Laboratory of Pharmacology and Toxicology
Graduate School of Pharmaceutical Sciences
Chiba University

Study focuses

1.

Identification of ribavirin uptake systems in human hepatocytes and characterization of their roles in ribavirin antiviral actions

In vitro ADME studies aiming at characterization of molecular mechanisms behind drug pharmacological or toxicological actions

2.

Identification of cancer-type OATP1B3 and its potential application to cancer therapy

3.

Establishment of new immortalized human brain cells for development of *in vitro* human BBB models

Study focuses

1.

Identification of ribavirin uptake systems in human hepatocytes and characterization of their roles in ribavirin antiviral actions

In vitro ADME studies aiming at characterization of molecular mechanisms behind drug pharmacological or toxicological actions

2.

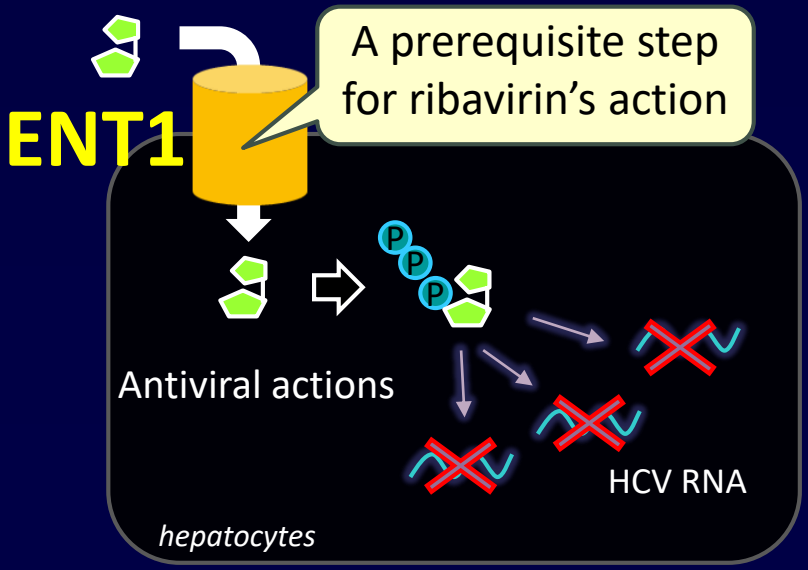
Identification of cancer-type OATP1B3 and its potential application to cancer therapy

3.

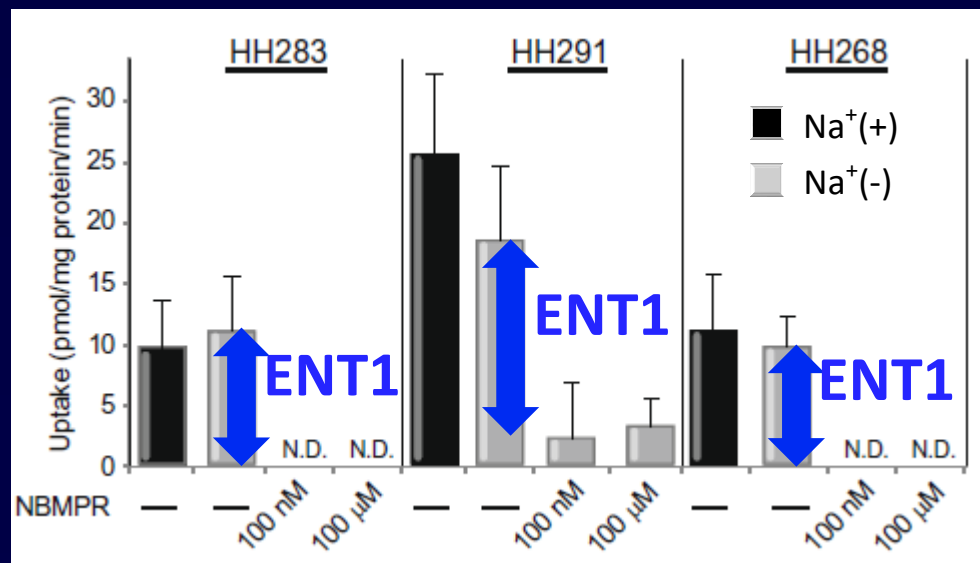
Establishment of new immortalized human brain cells for development of *in vitro* human BBB models

Identification of ribavirin uptake transporter in human hepatocytes

Ribavirin is a nucleoside analogue used for anti-hepatitis C virus (HCV) therapy (RBV)

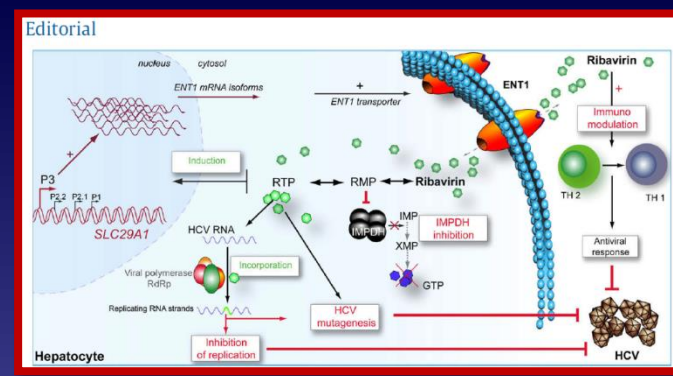
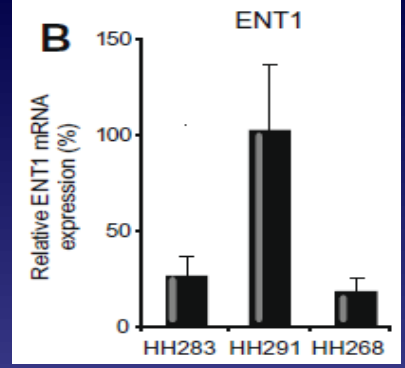


RBV uptake by human hepatocyte lines



Equilibrative nucleoside transporter 1 (ENT1) is a primary RBV uptake transporter in human hepatocytes.

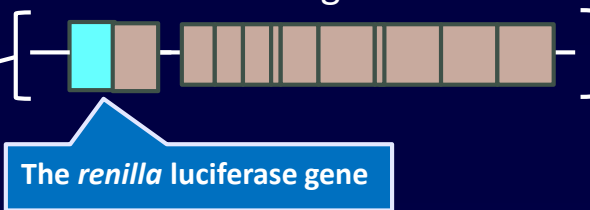
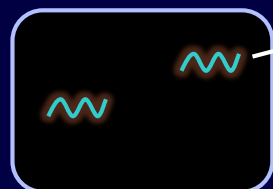
ENT1 mRNA level



Critical role of ENT1 in antiviral action of RBV in HCV-model cells

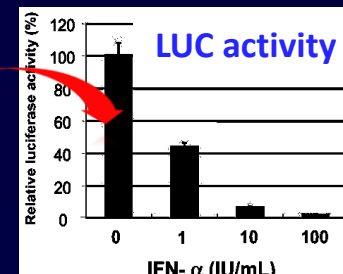
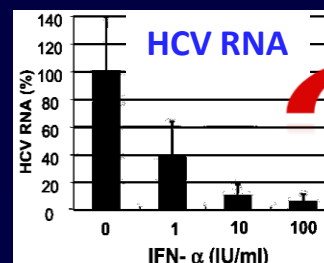
HCV-replication model cells (OR6 cells)

HCV genome



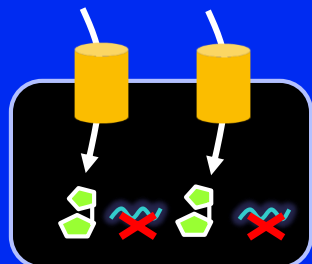
The *renilla* luciferase gene

The renilla luciferase activity level correlates well with the HCV replication activity level in OR6 cells



Ikeda, et al. *Biochem Biophys Res Commun* 2005;329:1350-9.

No ENT1 inhibition



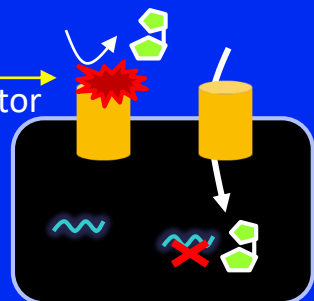
RBV uptake **100%**

HCV replication model cells

Concentration-dependent RBV activity (EC_{50})

203 ± 47 (μM)

ENT inhibitor



RBV uptake **50%**

399 ± 22 (μM)

0 100 200 300 400

RBV EC_{50} (μM)

Partial ENT1 inhibition

Ikura, Furihata, et al. *Antimicrob Agent Chemother* 2012;56:1407-13.

Association of an SNP in the *ENT1* gene with clinical outcome

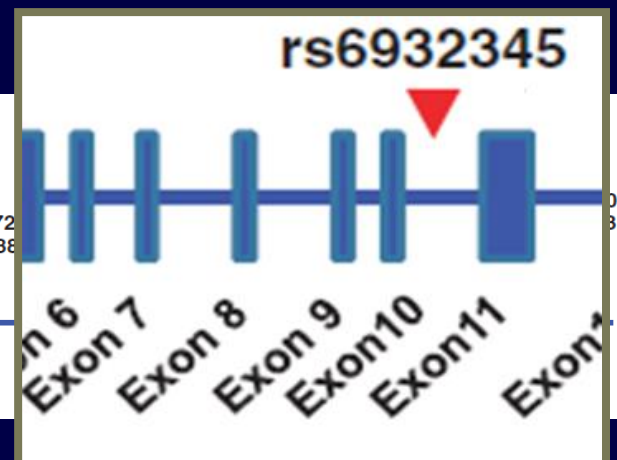
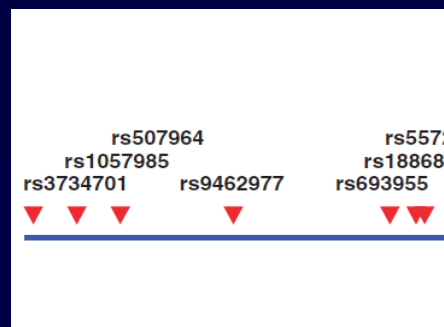
Collaborative work with Dr. Tsubota

Baseline profile of the patients recruited for the association study

Number (race): 526 (517 Japanese, 4/3/2 Chinese/Mongol/Korean)
HCV genotype: 1b
Regimen: Peg-INF/RBV combination therapy

SNPs in the *ENT1* gene analyzed in this study

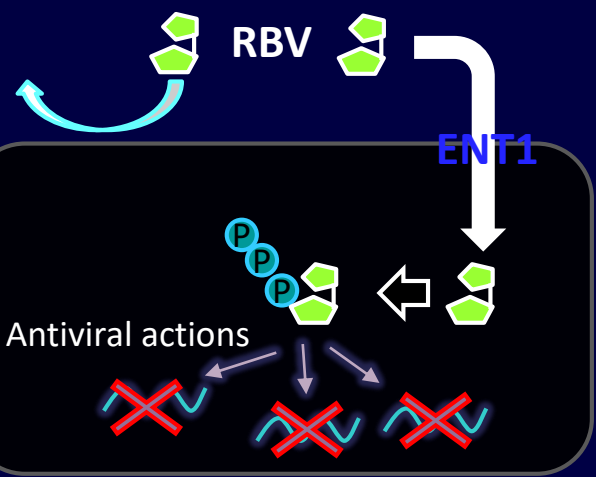
14 SNPs located in the 5'-upstream, intronic, or 3'-untranslated regions (indicated by red arrowheads)



Association of SNP rs6932345 with the rate of the sustained virological response (SVR, the therapeutic goal)

	Genotype (MAF)	<i>P</i> value on multivariate analysis	Odds ratio (95% CI)
rs6932345	AA vs AC/CC (0.196)	0.03	1.85 (1.06-3.21)

Summary & Perspective -1-

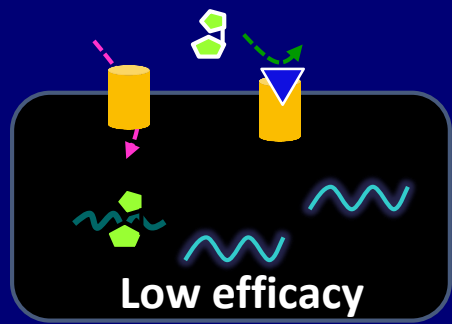


hepatocytes

ENT1 is a primary RBV uptake transporter in human hepatocytes. Accordingly, ENT1 plays a critical role in RBV's antiviral action.

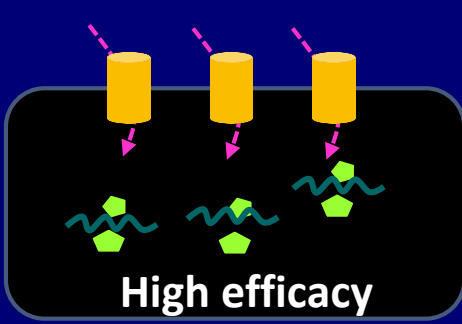
It can be assumed that hepatic ENT1 activity level is a factor that determines treatment efficacy of RBV-based anti-HCV therapy.

ENT1 inhibition by co-administered drug



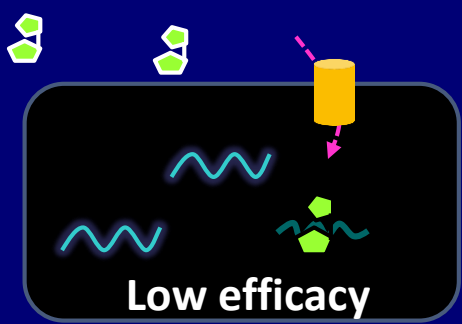
Low efficacy

Higher ENT1 expression /activity level ?



High efficacy

Lower ENT1 expression /activity level ?



Low efficacy

Study focuses

1.

Identification of ribavirin uptake systems in human hepatocytes and characterization of their roles in ribavirin antiviral actions

In vitro ADME studies aiming at characterization of molecular mechanisms behind drug pharmacological or toxicological actions

2.

Identification of cancer-type OATP1B3 and its potential application to cancer therapy

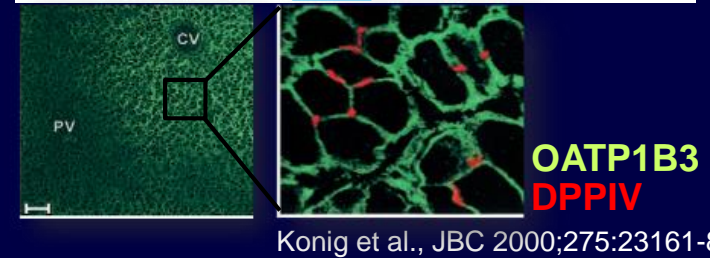
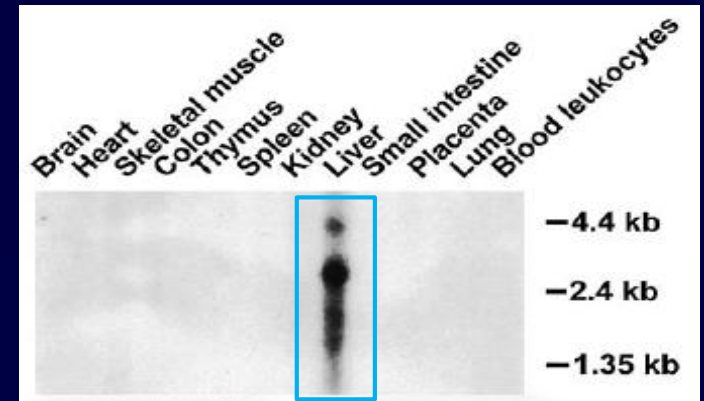
3.

Establishment of new immortalized human brain cells for development of *in vitro* human BBB models

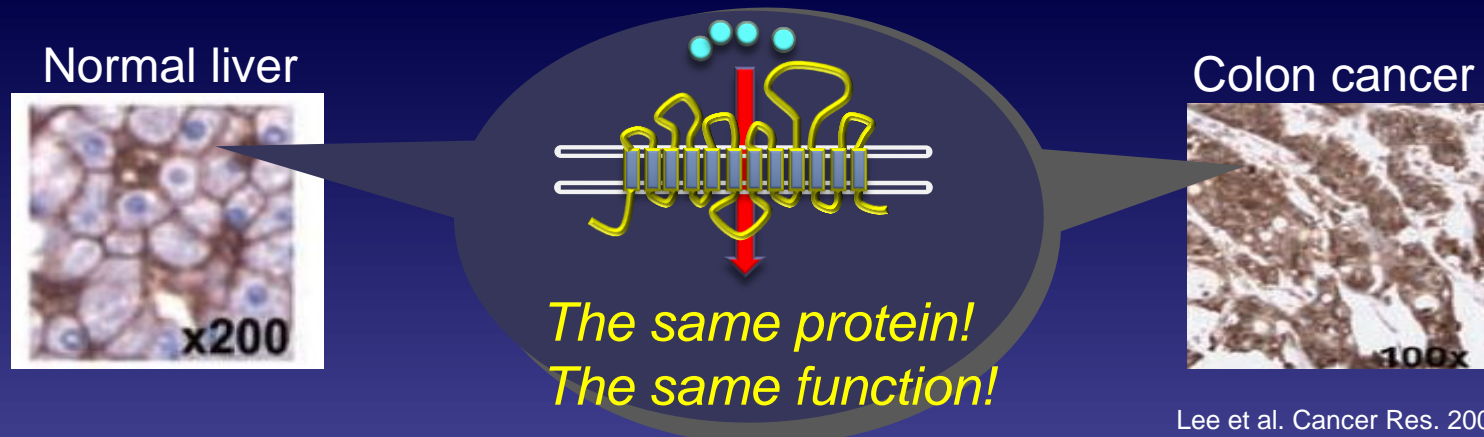
Organic anion transporting polypeptide 1B3 (OATP1B3)

OATP1B3

- ✓ is initially identified as a liver-specific transporter expressed at hepatocyte sinusoidal membrane.
- ✓ can transport various drugs into hepatocytes.
- ✓ is subsequently reported to be expressed in various cancer tissues.

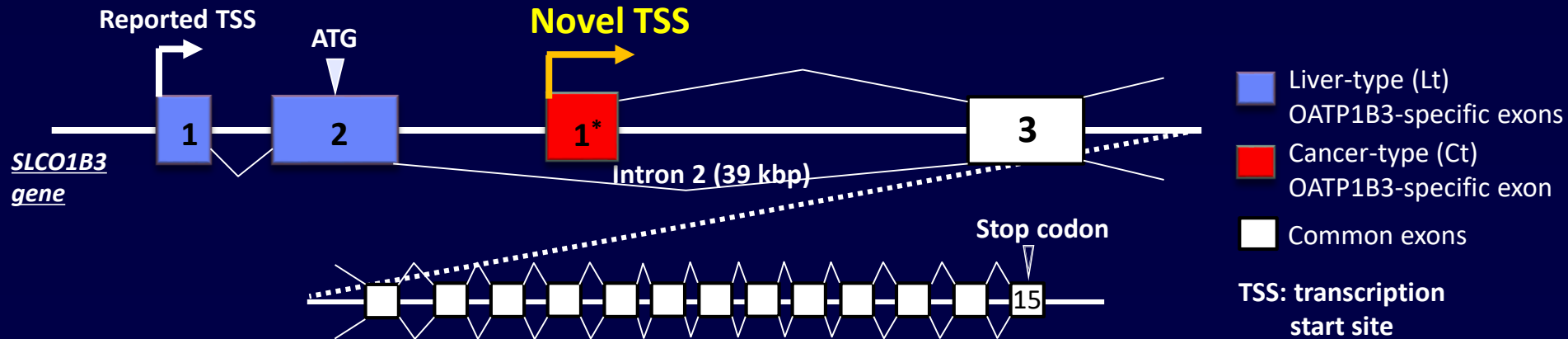
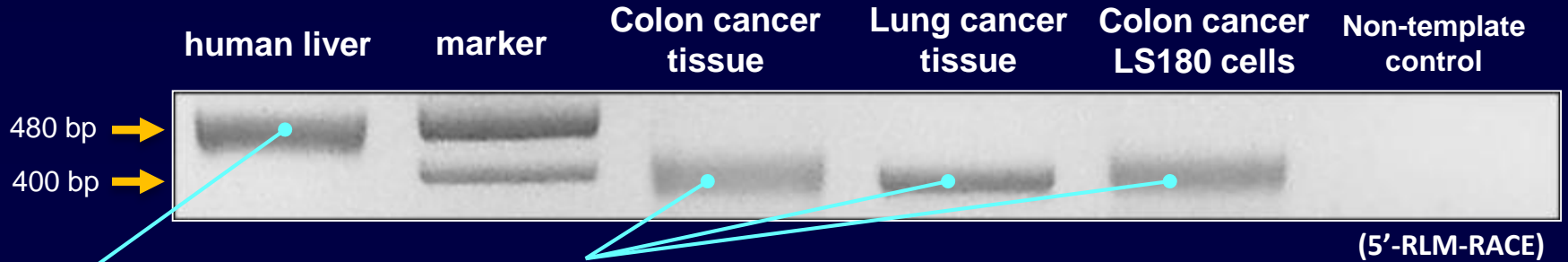


It had long been taken for granted that OATP1B3 expressed in cancer tissues was identical to that expressed in the liver.



Identification of cancer-type OATP1B3 in human cancer tissues

Cancer-type OATP1B3 (Ct-OATP1B3)



The new isoform identified in cancer is hereafter referred to as

Cancer type (Ct)-OATP1B3 mRNA



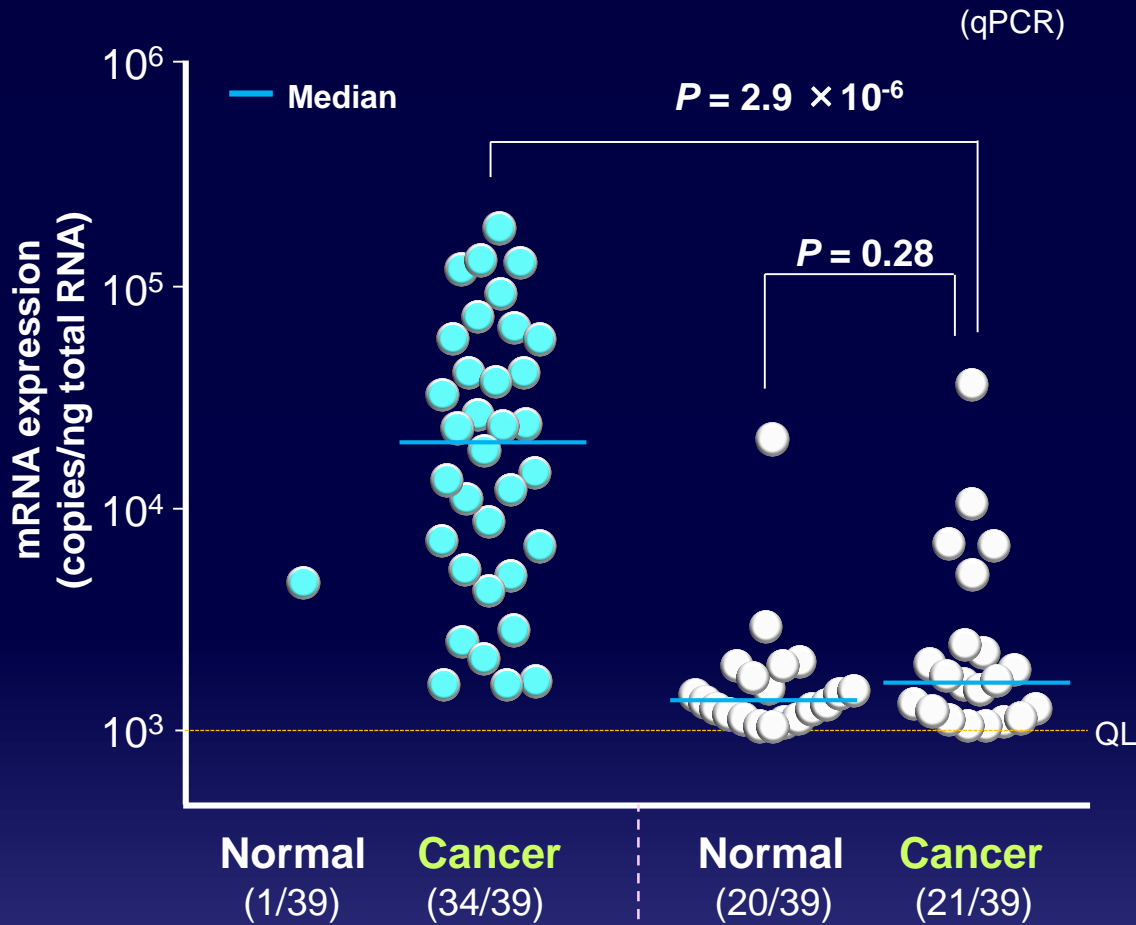
The known isoform identified in the liver is hereafter referred to as

Liver type (Lt)-OATP1B3 mRNA



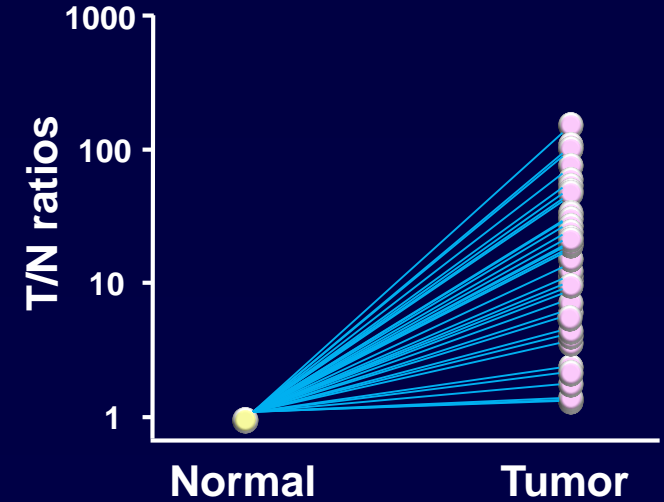
Cancer-restricted expression profile of ct-OATP1B3

OATP1B3 mRNA expression profile in 39 matched-pairs of colon tissues

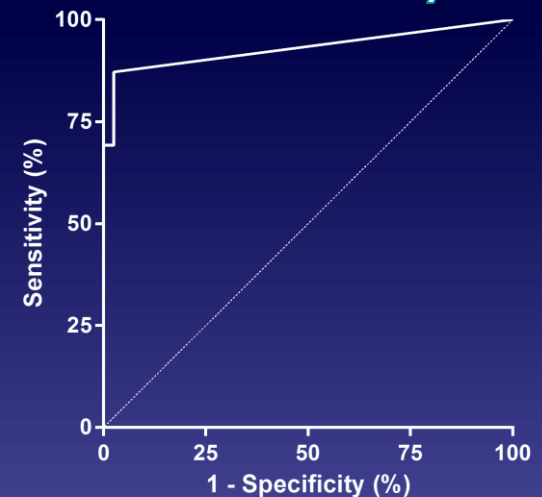


Ct-OATP1B3 | **Lt-OATP1B3**

T/N ratios of Ct-OATP1B3 mRNA levels in individual patients



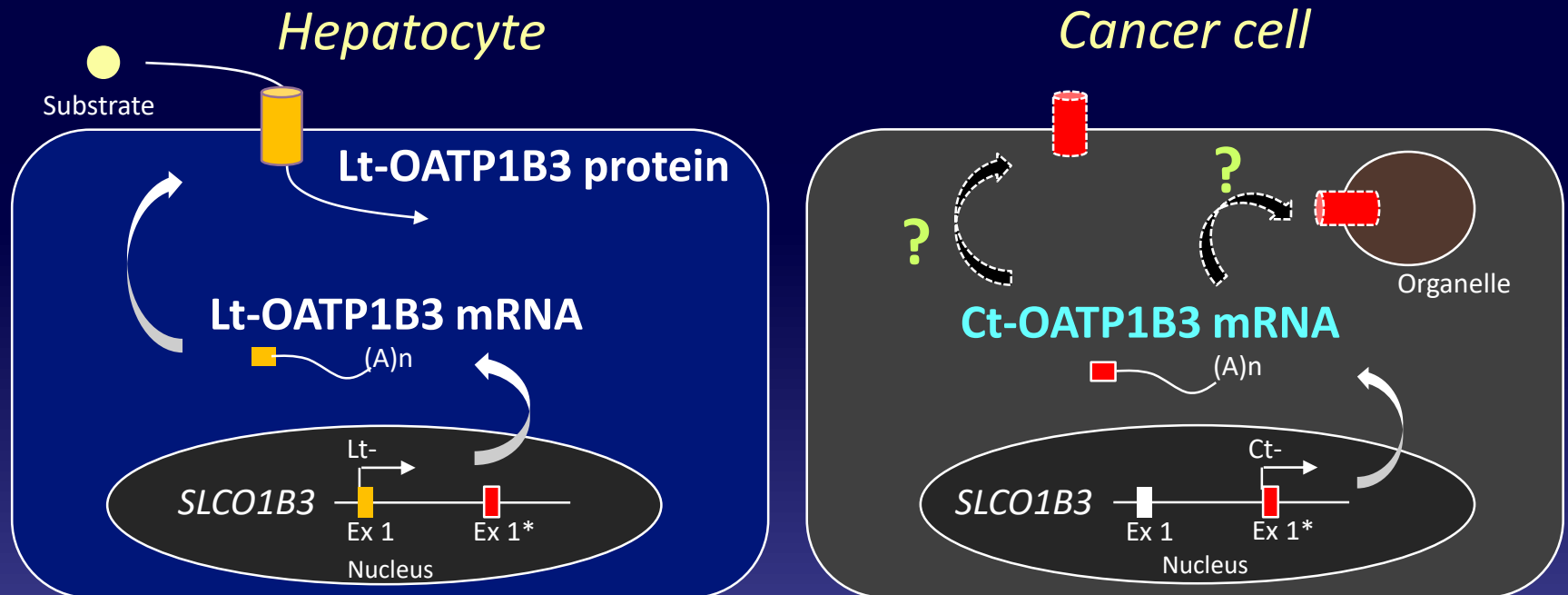
ROC curve analysis for evaluation cancer specificity and selectivity



Summary & Perspective -2-

Ct-OATP1B3, which is a variant isoform of Lt-OATP1B3, is considered to be a *bona fide* cancer-associated OATP1B3 isoform.

Identification of Ct-OATP1B3 is likely to revise the long-standing study premise, which is thus expected to open up new avenues in cancer-related OATP1B3 studies.



Study focuses

1.

Identification of ribavirin uptake systems in human hepatocytes and characterization of their roles in ribavirin antiviral actions

In vitro ADME studies aiming at characterization of molecular mechanisms behind drug pharmacological or toxicological actions

2.

Identification of cancer-type OATP1B3 and its potential application to cancer therapy

3.

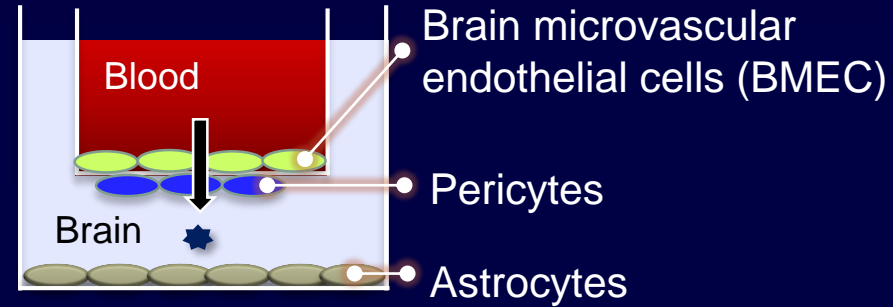
Establishment of new immortalized human brain cells for development of *in vitro* human BBB models

Blood-brain barrier and temperature-sensitive immortalized cells

Blood-brain barrier (BBB): A primary obstacle to drug penetration into the brain

In vitro BBB models can be applied to the different stages of CNS drug development.

In vitro BBB model



Primary human cells

are highly functional, but they show limited proliferation ability, rapid senescence, scarcity, and lot-to-lot variations, which

impede



Immortalized human cells

generally show infinite proliferation ability, human gene functions, stable phenotype, and cell-type specific functionality, which

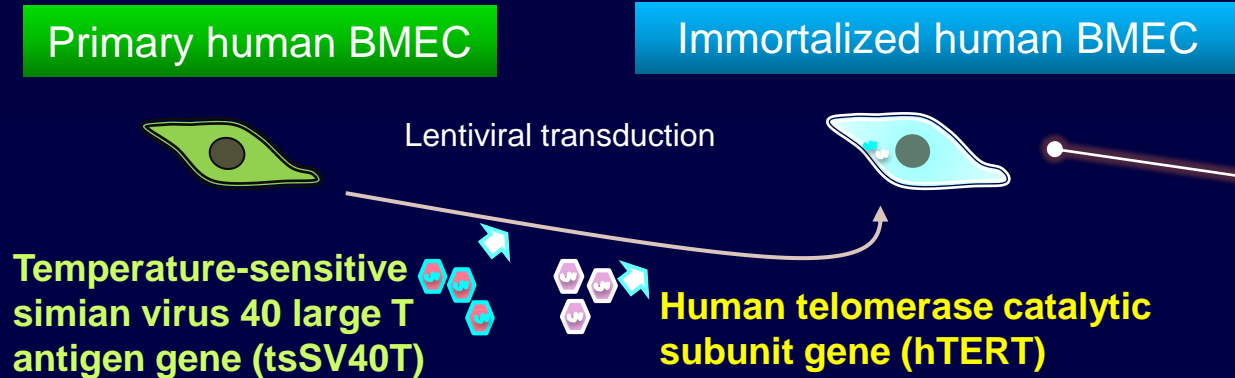


allow

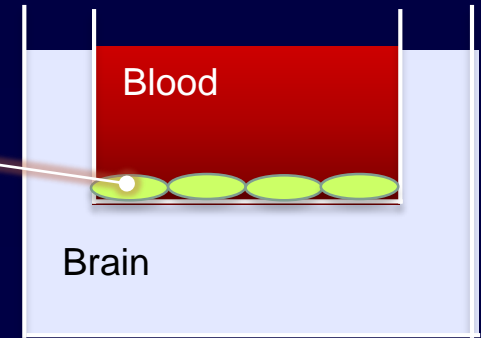
researchers to take various trial-and-error approaches in drug development.

Establishment of immortalized human BMEC

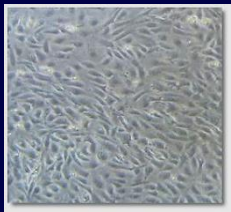
Establishment of human BMEC/conditionally immortalized clone β (HBMEC/ci β)



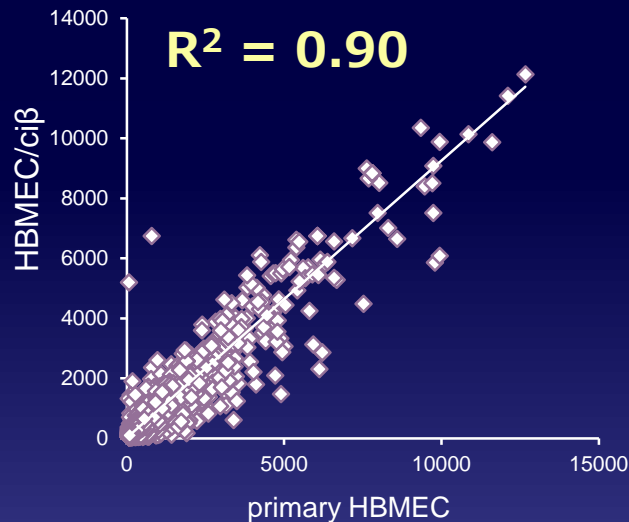
In vitro BBB model



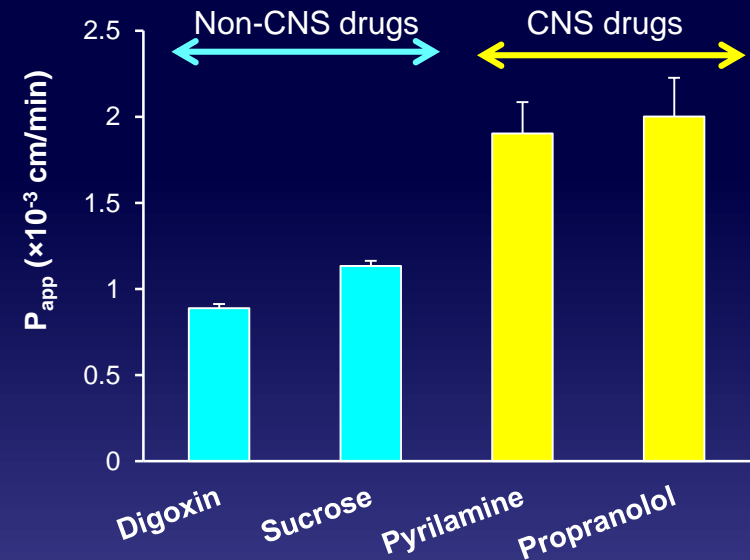
Morphology



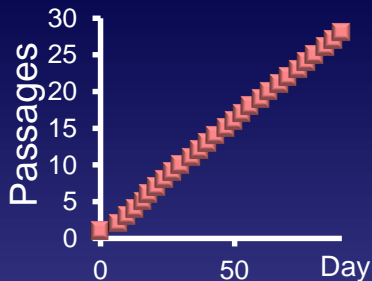
Global mRNA expression profile



Permeability analysis



Successive passages

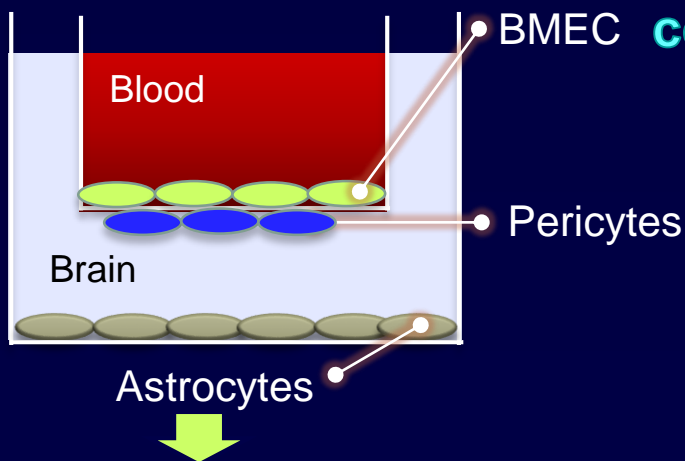


Kamiichi, Furihata et al. *Brain Res* 2012;1488:113-22
Furihata et al. *Fluids Barriers CNS*. 2015;5;12:7.

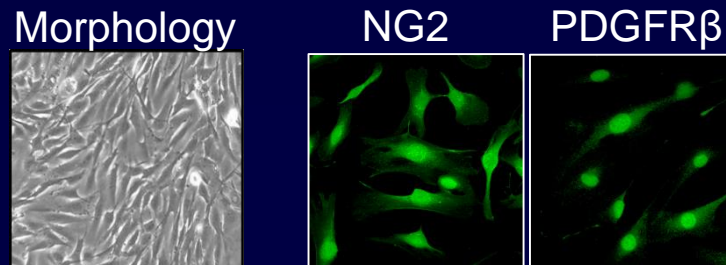
Furihata et al. unpublished.

Establishment of immortalized human astrocytes and pericytes

In vitro BBB model

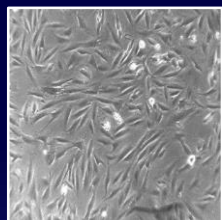


Establishment of human brain pericytes/conditionally immortalized clone 37 (HBPC/ci37)

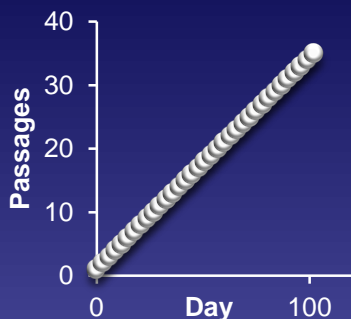


Furihata et al. unpublished.

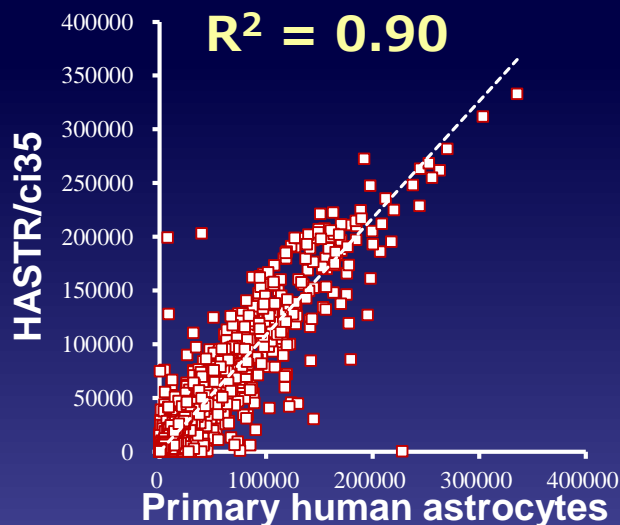
Establishment of human astrocyte/conditionally immortalized clone 35 (HASTR/ci35)



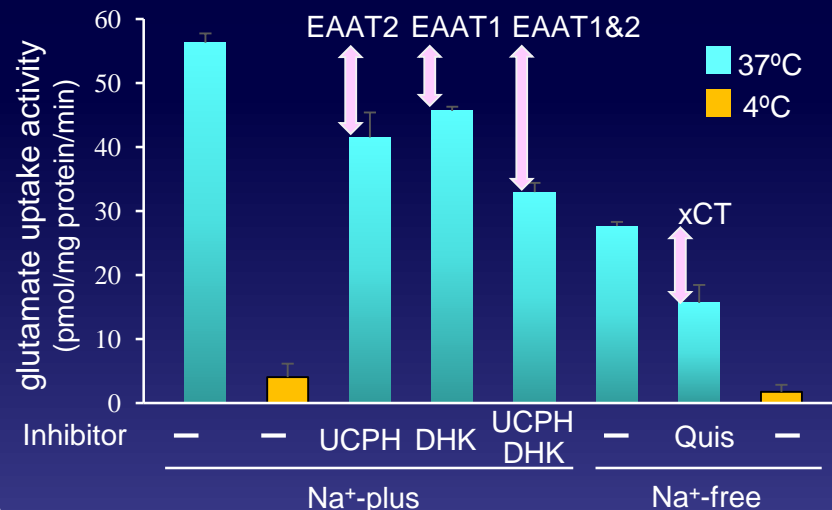
Successive passages



Global mRNA expression profile



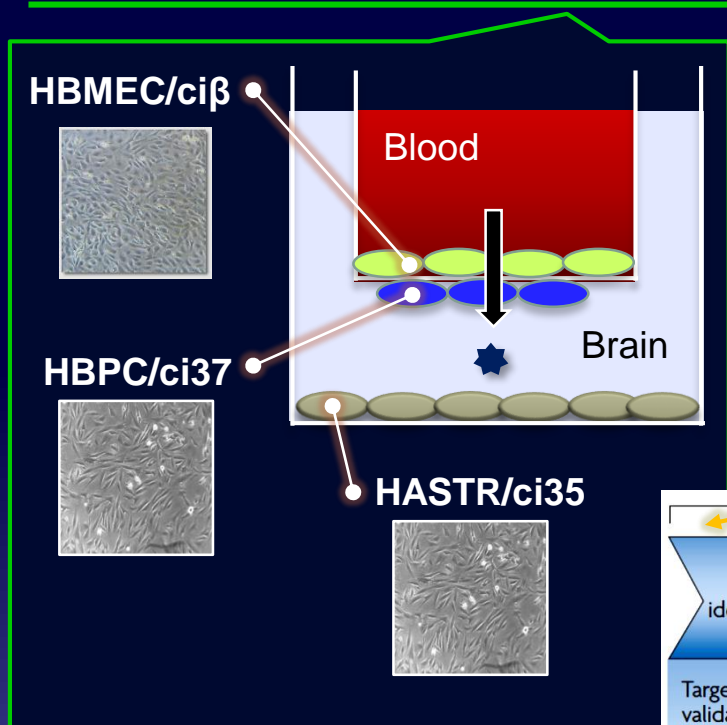
Glutamate uptake activity



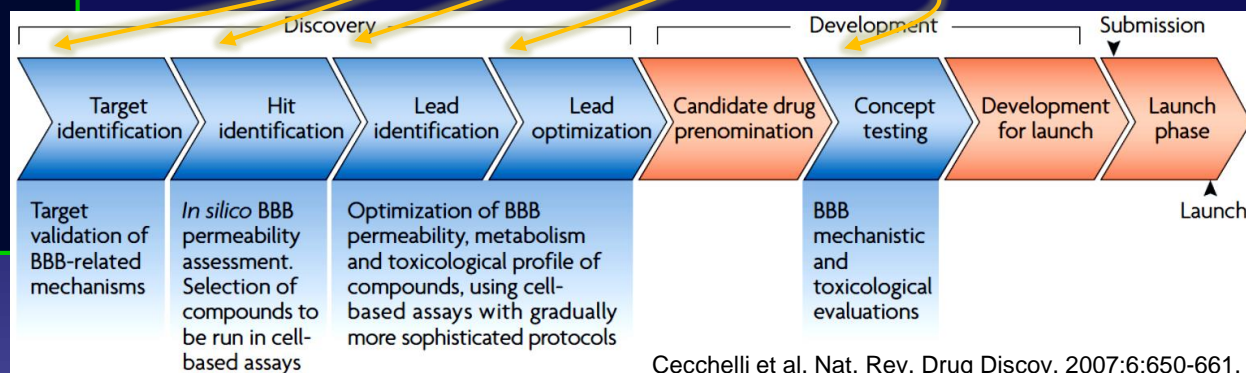
Furihata et al. J Neurochem. 2015, in press.

Summary & Perspective -3-

We have established HBMEC/ci β , which are highly proliferative and possess BBB properties. Furthermore, we have also established HASTR/ci35 and HBPC/ci37, as part of our ongoing efforts to develop an immortalized cell-based tri-culture *in vitro* BBB model.



Such models will be expected to provide easy-to-use, scalable, versatile, simplified working system in CNS drug development.



Acknowledgements

Laboratory of Pharmacology & Toxicology,
Chiba University

Dr. Kan Chiba

Dr. Kaoru Kobayashi

All laboratory members



Laboratory of Drug Metabolism &
Biopharmaceutics, Chiba Institute of Science

Dr. Masakiyo Hosokawa

Core Research Facilities for Basic Science,
Research Center for Medical Sciences, Jikei
University School of Medicine

Dr. Akihito Tsubota

Division of Medicinal Safety Science,
National Institute of Health Sciences

Dr. Yoshiro Saito

Dr. Kosuke Saito

Division of Pathology, Chiba Cancer
Centre Research Institute

Dr. Osamu Shimozato

Department of Medical Immunology, Graduate
School of Medicine, Chiba University

Dr. Shinichiro Motohashi

Laboratory of Biochemistry, Chiba University

Dr. Motoyuki Ito

Department of Tumor Virology, Okayama University
Graduate School of Medicine, Dentistry, and
Pharmaceutical Sciences

Dr. Nobuyuki Kato

Dr. Masanori Ikeda

Research Institute for Clinical Oncology,
Saitama Cancer Center

Dr. Takehiko Kamijo

Department of Thoracic Surgery, Graduate
School of Medicine, Chiba University

Dr. Ichiro Yoshino